

Review

Nitrogen Cycling and Mass Balance in the World's Mangrove Forests

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Abstract: Nitrogen (N) cycling in mangroves is complex, with rapid turnover of low dissolved N concentrations, but slow turnover of particulate N. Most N is stored in soils. The largest sources of N are nearly equal amounts of mangrove and benthic microalgal primary production. Dissolved N fluxes between the forests and tidal waters show net uptake, indicating N conservation. N_2 -fixation is underestimated as rapid rates measured on tree stems, aboveground roots and cyanobacterial mats cannot currently be accounted for at the whole-forest scale due to their extreme patchiness and the inability to extrapolate beyond a localized area. Net immobilization of NH_4^+ is the largest ecosystem flux, indicating N retention. Denitrification is the largest loss of N, equating to 35% of total N input. Burial equates to about 29% of total inputs and is the second largest loss of N. Total inputs slightly exceed total outputs, currently suggesting net N balance in mangroves. Mangrove PON export equates to $\approx 95\%$ of PON export from the world's tropical rivers, but only 1.5% of the entire world's river discharge. Mangrove N_2O emissions, denitrification, and burial contribute 0.4%, 0.5–2.0% and 6%, respectively, to the global coastal ocean, which are disproportionate to their small worldwide area.

Keywords: biogeochemistry; denitrification; ecosystem; mangrove forests; mass balance; nitrogen; nitrogen cycling; nitrogen fixation; nitrogen retention

1. Introduction

Nitrogen (N) is the most important nutrient element in fostering growth, reproduction, productivity, and other energetic and physiological processes that enable ecosystems to thrive. Mangrove forests and their associated waterways are major coastal ecosystems that live along the world's subtropical and tropical coastlines, requiring nitrogen and other nutrients like all other ecosystems [1]. Mangroves are an important ecological and economic resource, offering a wide variety of ecosystem goods and services, such as being important breeding sites and nursery grounds for birds, fish, crustaceans, amphibians, shellfish, reptiles, and mammals [2]. These tidal forests are a potentially renewable resource of wood and accumulate sediment, carbon, contaminants and nutrients, such as N. Mangroves also provide a vital livelihood for coastal inhabitants and offer some protection against coastal erosion and catastrophic events, such as tsunamis [1,2]. Thus, it is important to understand how these tropical ecosystems function biogeochemically.

In mangrove ecosystems where N is often limiting, native flora and fauna and their associated food webs have evolved a variety of mechanisms to conserve N. These retention mechanisms and strategies include, but are not restricted to: (1) highly efficient solute uptake between trees, microbes and soil N pools; (2) high N-use efficiency and high rates of leaf resorption; (3) low rates of N loss, such as dissolved N export and nitrous oxide (N_2O) emissions in proportion to N inputs; (4) export of highly refractory N in the form of humic and fulvic acids (tannins); (5) rapid rates of nitrogen fixation at the soil surface and on various forest components (bark, downed wood, prop roots, pneumatophores, cyanobacterial mats); and (6) a large reservoir of dead roots belowground [1,2].

Living in a low-N environment, mangroves rely greatly on the efficiency of the microbial machinery existing in their soils and tidal waters to process and conserve N in its various forms. Numbers, diversity and productivity of soil and planktonic bacterial communities are high, resulting in rapid uptake, transformation, and release of particulate and dissolved inorganic and organic N [1,3]; dissolved N pools turn over rapidly, usually in a matter of minutes to days [4]. The experimental addition of nutrients to mangrove trees in the field and laboratory indicate rapid uptake and utilization, although complex patterns often result due to various interactive factors, such as differences in forest stands in species composition, soil type (carbonate sand, terrigenous silt/clay, quartz sand, etc.), intertidal position, soil fertility, salinity, forest age, and tree and forest development stage [5–7].

Despite our knowledge of N transformation and utilization processes and rates in mangrove ecosystems, a comprehensive picture at the ecosystem-level has yet to emerge. To this day, only one complete N budget exists for a mangrove ecosystem [4] and no attempt has been made to construct a N mass balance for the world's mangrove ecosystems. This paper details such an attempt as a model tool to identify what is and what is not known about N cycling processes in mangroves. First, we will assess the standing stocks of N in the forests and associated waterways, prior to examining functional processes, including the significance of N retention and the contribution of mangroves to N flow in the global coastal ocean.

2. Nitrogen Concentrations and Standing Stocks

2.1. Dissolved N Concentrations in Tidal Waters and Porewater

Dissolved organic and inorganic N in mangrove tidal waters are dominated by dissolved organic nitrogen (DON), followed in descending order by ammonium (NH_4^+), nitrate (NO_3^-) and nitrite (NO_2^-). In nearly all cases, except for some organically polluted environments, concentrations of all four species are in the micromolar (μM) range. DON, NH_4^+ , NO_2^- and NO_3^- concentrations in unpolluted waterways range from 0.1 to 60 μM , 0 to 120 μM , 0 to 5 μM and 0 to 37 μM , respectively [3,4]. The major drivers of change in concentrations are rainfall, land runoff, intrusions of offshore water, groundwater and porewater inputs, leaching from litter, temperature, anthropogenic inputs, and plankton metabolic activities, especially of phytoplankton and bacterioplankton [1].

In soil porewater, dissolved N concentrations also vary greatly, depending on soil particulate N concentrations and composition, soil texture, redox status, presence of sulfides and other reductants, microbial mineralization processes, salinity, bioturbation, rate of soil accumulation, benthic community composition, and, most significantly, uptake and release from mangrove roots [1]. The chemical composition of DON in tidal waters and in porewater is not well characterized, but some evidence indicates that most DON is composed of humic acids (range: 47–91%) and to a lesser extent, of aromatic compounds, peptides and amino acids [8].

Figure 1 illustrates the importance of tree uptake on NH_4^+ porewater concentrations, showing an inverse relationship between the vertical soil depth changes in porewater NH_4^+ concentrations and live fine root biomass in three *Kandelia candel* forests in southern China [9]. A Pearson product-moment correlation (r) of porewater ammonium versus root biomass of -0.476 is significant ($p < 0.00784$). Earlier data [10,11] have similarly indicated an effect of NH_4^+ uptake by mangrove roots on porewater concentrations and profiles. Additional evidence indicates that adjacent mudflat sediments usually have higher porewater concentrations of NH_4^+ than in mangrove soils, suggesting that the difference is due to tree uptake [11]. These data are circumstantial, but support experimental data showing a strong preference by mangroves for ammonium (and nitrate) for their growth and nutrition [12–15]. Nitrite and nitrate are often found in mangrove porewater [4,11], but concentrations show very irregular patterns with increasing soil depth.

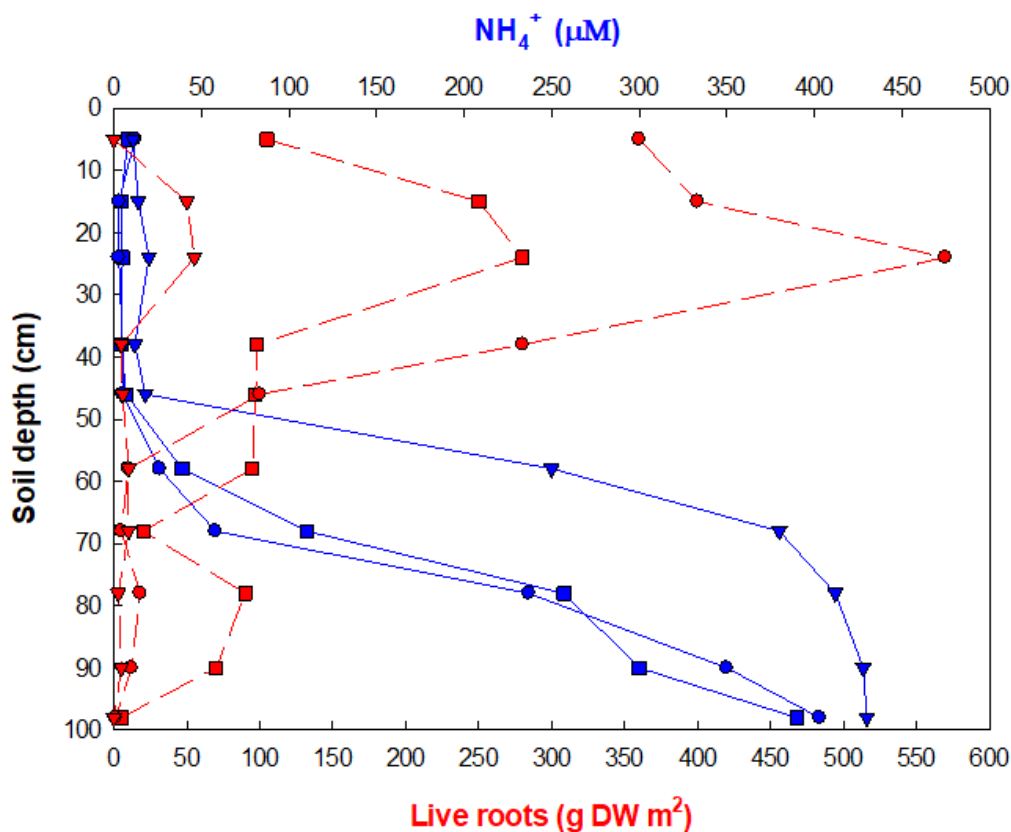


Figure 1. Vertical profiles of porewater extractable ammonium (NH_4^+ , μM) compared with live root biomass (g dry weight m^2) in three *Kandelia candel* forests in the Jiulongjiang estuary, China [9].

2.2. Total N Concentrations and C/N Ratios in Forest Components and Soil

Concentrations of total nitrogen (TN) in leaves, stems, branches, roots, litter, and soil vary among forests due to many factors, including species composition, forest age, allochthonous N inputs, extent of organic pollution (if any), climate, temperature, salinity, physiological condition, nutritional status, soil composition, porewater nutrient concentrations, micronutrient availability, rate of soil organic matter accumulation, intertidal position, tidal regime, redox status, presence of sulfides and other reductants, microbial mineralization processes, bioturbation, and benthic community composition [1,4]. On average, TN concentrations in mangrove forests are very similar to those in tropical terrestrial forests for leaves and wood, and lower (Table 1) than terrestrial forest soil N (mean = 0.45% DW) and roots (mean = 0.86% DW), but higher (Table 1) than the C/N ratio (g/g) of below-ground biomass (mean = 33.2) (terrestrial forest references [16–38]).

Table 1. Mean (± 1 standard error, SE) and median total nitrogen (TN) concentrations (as percentage of dry weight, DW) of various above- and belowground mangrove components, including soils to a depth of 1 m. The mean C/N ratios (g/g) are as follows: soil C/N = 11.3; live wood C/N = 130.4; belowground root C/N = 80.9; litter C/N = 79.4. The C/N values for biomass were calculated based on relative percentage of biomass DW in the respective components (i.e., stems + branches for wood estimate) and dead and live root and rhizome DW for belowground estimates using the carbon data and references in [39]. N data from [4,9–11,39–112] and earlier references within.

Component	Number of Observations	Mean ± 1 SE	Median	Range
Green leaves (%N)	109	1.61 \pm 0.05	1.54	2.55
Stems (%N)	58	0.37 \pm 0.03	0.35	1.22
Branches (%N)	55	0.31 \pm 0.02	0.29	0.65
Roots (%N)	63	0.49 \pm 0.03	0.45	1.69
Litter (%N)	153	0.69 \pm 0.03	0.64	2.04

2.3. Global Mean N Stocks

Mean TN stock of global mangrove forests (Figure 2) totals 52.03 Mg N ha⁻¹, with 96% of total TN stock contained in soils to a depth of 1 m (mean = 50.0 Mg N ha⁻¹). In comparison, mean TN stocks in tropical terrestrial forests (rain forests, other moist forests, and peat swamps) total 22.33 Mg N ha⁻¹ (Figure 2). Like mangroves, 91% of N stocks are vested in soil (mean = 20.3 Mg N ha⁻¹), suggesting similar investment of N by both mangroves and terrestrial forests in various forest components and belowground storage.

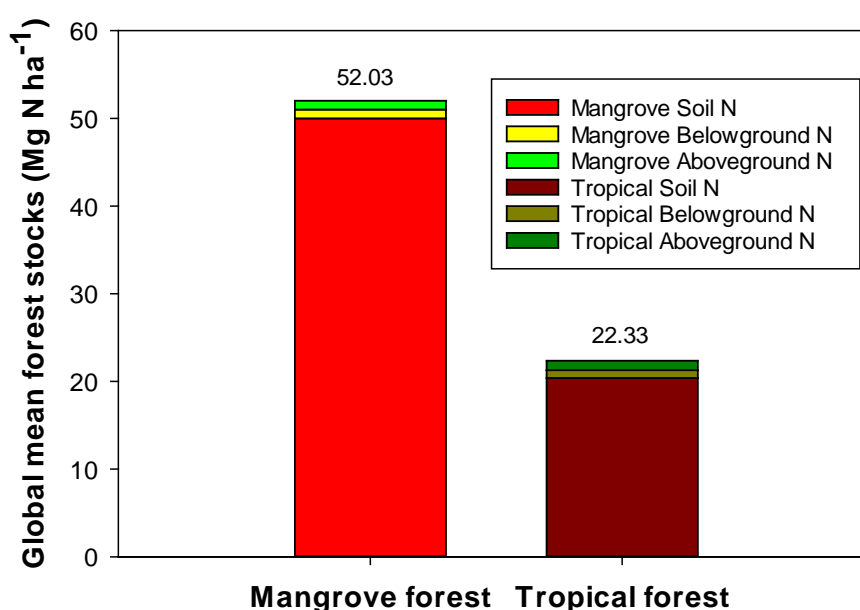


Figure 2. Mean TN stocks in mangrove above- and belowground biomass and soils to a depth of 1 m. Biomass and soil dry weight data extrapolated to N using N content or C/N ratios (g/g) in Table 1. Tropical terrestrial forest data are presented for comparison and includes data from moist forests, peat swamp forests, and rain forests with N, C and C/N (g/g) data [16–38,113–120] and earlier references within.

3. The Nitrogen Cycle in Soils

The soil nitrogen cycle (Figure 3) is composed of a series of complex transformation processes, nearly all conducted by specialized bacterial and archaeal groups, such as ammonium-oxidizers, cyanobacteria, nitrate-reducers, and nitrite-oxidizers.

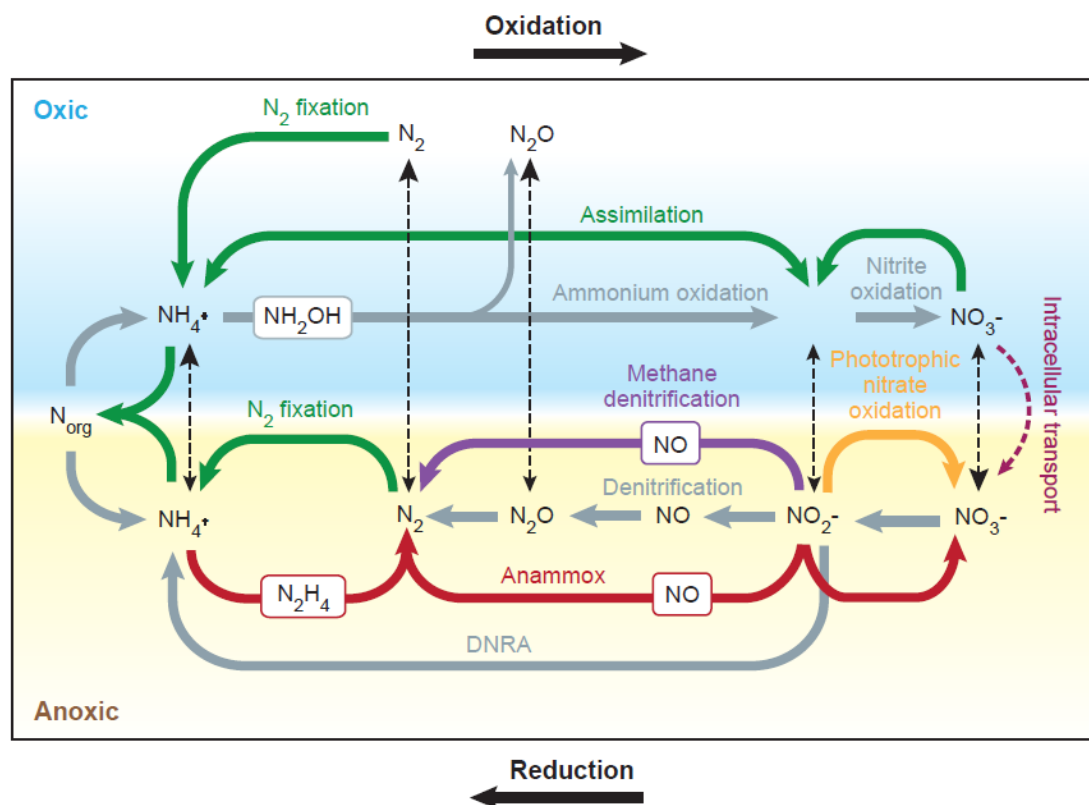


Figure 3. The nitrogen cycle showing all oxidizing and reducing transformation pathways that occur in oxidized and anoxic environments, particularly in marine sediments and waterlogged saline soils. Ammonification (shown in gray on left side) is the microbial breakdown of organic nitrogen (N_{ORG}) into ammonium (NH_4^+); the reverse process is NH_4^+ immobilization (green arrows on left). Nitrification (gray arrows in top side) is the biological oxidation of NH_4^+ to NO_2^- followed by the oxidation of the NO_2^- to NO_3^- . Black dotted arrows depict transport processes between oxic and anoxic environments. Abbreviations: DNRA = dissimilatory nitrate reduction to ammonium; Anammox = anaerobic ammonium oxidation; N_2 fixation = nitrogen fixation; NO = nitric oxide; N_2O = nitrous oxide; N_2 = nitrogen gas; NH_2OH = hydroxylamine, an intermediate in biological nitrification; N_2H_4 = hydrazine, the intermediate in the anaerobic oxidation of ammonium (anammox) process. Reproduced with permission from [121].

Fungi are also involved in organic matter decomposition and in carbon and nitrogen cycling processes, especially on litter and in roots and rhizomes [122–124]. In mangrove soils, a vast menagerie of highly diverse bacterial, archaeal, fungal and protistan communities exist, comprised mostly of members of the phyla Proteobacteria and Bacteroidetes, Bathyarchaeota and Euryarchaeota, Ascomycota, and Sarcondina, Mastigophora, Ciliophora and Myxomycota. Interkingdom biotic factors shape both community structure and function of all four kingdoms. Aside from fully oxic and anoxic conditions, mangrove soils provide microaerophilic environments, such as in and near biogenic structures and in rhizospheres where microaerophiles carry out nitrogen reactions such as ammonium oxidation [125]. All of these microbial types and communities vary greatly in composition and function in relation to such factors as temperature, mangrove species composition, tides, pH, salinity, and soil fertility, redox status, and soil type [126,127].

Several microbial groups and transformation processes have only recently been discovered (DNRA, anammox, methane denitrification), but are often quantitatively important in N cycling. For example, a nitrite-dependent anaerobic methane-oxidizing bacterium was recently discovered in mangrove soils of the Zhangjiang estuary in China [128]. This "*Candidatus Methylomirabilis*

oxyfera-like" bacterium is unique in linking the carbon and nitrogen cycles. It is conceivable that new microbe species and new N transformation pathways await discovery in mangrove environments.

At the ecosystem-scale, other N processes are important in mangroves, such as the exchange of dissolved and particulate nitrogen (DN, PN) between the forests and tidal waterways and with the adjacent coastal ocean, dry and wet deposition of N, groundwater inputs, sedimentation and burial, and assimilation and retention of N by flora and fauna.

3.1. Forest Soil N Transformations

3.1.1. Nitrogen Fixation

N₂-fixation rates are highest on algal/bacterial crusts growing on tree stems (Table 2), although the few measurements preclude meaningful statistical analyses. One-way analysis of variance on ranks ($H = 46.082$; $p < 0.001$) followed by Dunn's method (Q) indicate significant differences between rates measured on the soil surface and the other forest vegetation. Rates of N₂-fixation on pneumatophores and prop roots are significantly greater than on the soil surface and vegetation components (Q values = 3.757–4.743; $p < 0.001$), except for belowground roots and rhizomes. Although few measurements have been made on vegetation and cyanobacterial mats, N₂-fixing microbial communities associated with roots, stem surfaces, and cyanobacterial mats may provide significant input of N to mangrove forests.

Table 2. Rates of nitrogen fixation ($\text{mg N m}^{-2} \text{ d}^{-1}$) at the mangrove soil surface, cyanobacteria mats, aboveground roots (pneumatophores and prop roots), belowground roots and rhizomes, litter and senescent leaves lying on the forest floor, and microbial crusts on the bark of tree stems [1,4,9,55–58, 129–161] and earlier references within. Rates presented on a $\text{g}^{-1} \text{ DW/WW}$ basis were converted to m^{-2} assuming $1 \text{ g matter} = 1 \text{ cm}^{-3}$. Ethylene rates were converted to N using the theoretical factor of $3 \text{ C}_2\text{H}_2 \text{ molecules} = 1 \text{ N molecule}$ [162].

Component	Number of Observations	Mean \pm 1 SE	Median	Range
Soil surface	57	8.22 ± 1.69	3.22	0–58.92
Cyanobacteria mats	28	9.69 ± 2.62	3.43	0–60.42
Aboveground roots	18	31.78 ± 5.63	26.25	2.77–73.4
Belowground roots + rhizomes	15	6.21 ± 1.65	4.50	1.04–26.20
Litter	9	1.16 ± 0.40	0.45	0.21–3.30
Senescent leaves	7	0.75 ± 0.13	0.83	0.39–1.20
Stem bark	5	100.95 ± 36.42	100.95	17.40–201.20

3.1.2. Within-Soil Transformations

Denitrification has been measured often in mangrove soils, mostly within the upper 5–20 cm (only complete (to N₂ production) denitrification rates were considered, Table 3). Gross rates of soil ammonification are significantly greater than net ammonification rates (one-way ANOVA on ranks, $H = 33.483$; Dunn's $Q = 5.786$; $p < 0.001$). There are no significant differences between gross and net rates of soil nitrification, although mean rates of gross nitrification are nearly three times greater than net rates (Table 3), indicating significant N immobilization. Although some studies found that anammox is a minor transformation process in mangrove soils compared to complete denitrification (references in Table 3), the available data (Table 3) show no significant differences between rates of both processes (one-way ANOVA on ranks, $H = 0.788$; $p = 0.375$). Rates of microbial N metabolism depend on many drivers, including temperature, soil fertility, microbial community structure, plant metabolism and root activities, bioturbation, intertidal position, and salinity. There is evidence that other N-transforming bacterial and archaeal groups (see Section 3) are present in mangrove soils, but no rate data are available for processes, such as methane denitrification, nitrite oxidation and phototrophic nitrate oxidation.

Table 3. Rates of N transformation processes ($\text{mg N m}^{-2} \text{d}^{-1}$) in mangrove soils. All rates are to soil depths of 5–20 cm. Rates presented on a g^{-1} DW/WW basis were converted to m^{-2} assuming $1 \text{ g soil} = 1 \text{ cm}^{-3}$. Abbreviations: DNRA= dissimilatory nitrate reduction to ammonium; anammox = anaerobic ammonium oxidation. Source: [9,15,55–58,129,133–192].

Transformation Process	Number of Observations	Mean \pm 1 SE	Median	Range
Denitrification (complete)	165	26.25 ± 3.34	3.90	0–443.52
Gross ammonification	17	301.63 ± 50.90	267.40	77.40–898.80
Net Ammonification	52	31.40 ± 6.19	15.40	0.17–200.00
Gross nitrification	25	15.08 ± 5.81	4.74	0–141.00
Net nitrification	37	5.80 ± 1.20	1.93	0–30.80
Anammox	35	22.11 ± 5.49	4.35	0–99.4
DNRA	21	18.19 ± 6.77	4.54	0.01–108.64

3.1.3. NO , N_2O and DN Fluxes from Surface Soils

Only two measurements at one site have been made of NO fluxes across the soil–air interface (Table 4); both indicate negligible release. Rates of N_2O flux show a net mean flux to the atmosphere, although some measurements indicate net uptake by mangrove soils (Table 4).

Rates of DON , $\text{NO}_2^- + \text{NO}_3^-$, and NH_4^+ (Table 4) flux vary widely among forests, due mostly to differences between fluxes measured in light (clear) versus dark (opaque) chambers. Most measurements from light bottles show net uptake of solutes indicating utilization of dissolved nutrients in overlying tidal waters by microalgae, cyanobacteria, and other autotrophs on the mangrove soil surface. This fact is reflected in the mean rates of exchange being negative for all three dissolved N forms. Due to high variation, all three solutes appear to be utilized equally by benthic autotrophs.

Table 4. Estimates of net NO and N_2O gas and dissolved nitrogen fluxes across the mangrove soil surface. Units = $\text{mg N m}^{-2} \text{d}^{-1}$. Negative values indicate fluxes into the soil and positive values indicate fluxes from the soil to the overlying tidal water/ atmosphere. Source: [9,164,165,167,169,176,184,193–210].

Gas and Solute Soil Fluxes	Number of Observations	Mean \pm 1 SE	Median	Range
N_2O soil–air flux	69	0.60 ± 0.17	0.22	–2.03–9.01
NO soil–air flux	2	0.05 ± 0.009	0.05	0.04–0.06
DON soil–water flux	41	-18.29 ± 18.14	0.00	–743–19.6
$\text{NO}_2^- + \text{NO}_3^-$ soil–water flux	66	-4.46 ± 1.05	–0.24	–29.04–3.36
NH_4^+ soil–water flux	78	-1.48 ± 2.20	–0.16	–88.7–55.19

4. Tidal Water N Processes

4.1. N_2O Fluxes

Fluxes of N_2O in mangrove tidal waters have been measured in more than 30 mangrove-fringed estuaries, tidal creeks, and waterways for a total of 62 measurements [211–226]. Net flux is to the atmosphere, averaging (± 1 SE) $0.11 \pm 0.03 \text{ mg N m}^{-2} \text{d}^{-1}$ with a median of $0.02 \text{ mg N m}^{-2} \text{d}^{-1}$ and ranging from net uptake ($-0.06 \text{ mg N m}^{-2} \text{d}^{-1}$) to net release ($1.32 \text{ mg N m}^{-2} \text{d}^{-1}$).

4.2. Tidal Exchange

Rates of exchange between mangrove forests and adjacent tidal waterways vary greatly within and among sites, as reflected in the wide range of estimates for NH_4^+ , $\text{NO}_2^- + \text{NO}_3^-$ and DON flux (Table 5). Despite the variability, net exchange is into the mangroves for all DN species based on mean rates; median rates indicate little net exchange (Table 5). The behavior of DN in transport between tidal waters and mangroves depends on a wide range of factors, including tidal prism, geomorphology, climate, seasonal weather patterns, the ratio of forest to waterway area, temperature, salinity, pH, dissolved oxygen levels, and plankton metabolism [1,3,4].

Table 5. Rates of NH_4^+ , DON and $\text{NO}_2^- + \text{NO}_3^-$ exchange between mangrove forests and adjacent tidal waterways, including mangrove-fringed estuaries. Source: [1,4,227–241] and references within. Import into the mangroves is shown as a negative value and export to adjacent tidal waters is a positive value. Units = $\text{mg N m}^{-2} \text{ water d}^{-1}$.

Component	Number of Observations	Mean \pm 1 SE	Median	Range
NH_4^+ import	26	-9.81 ± 4.35	-1.57	-100.76--0.006
NH_4^+ export	23	8.00 ± 3.63	0.39	0.03–62.40
NH_4^+ net exchange	49	-1.45 ± 3.12	-0.01	-100.76–62.4
DON import	9	-4.69 ± 2.17	-2.87	-21.84--0.91
DON export	10	1.45 ± 0.33	1.37	0.08–3.27
DON net exchange	19	-1.46 ± 1.24	0.08	-21.84–3.27
$\text{NO}_2^- + \text{NO}_3^-$ import	24	-3.62 ± 1.52	-0.73	-28.80--0.003
$\text{NO}_2^- + \text{NO}_3^-$ export	27	1.84 ± 0.55	0.29	0.05–11.90
$\text{NO}_2^- + \text{NO}_3^-$ net exchange	51	-0.73 ± 0.85	0.08	-28.8–11.90

5. An Ecosystem-Level View of Mangrove Forests: A N Budget

Using the data in the preceding tables, a preliminary N budget for the world's mangrove forests was constructed (Figure 4). The mean values used in the budget are not absolute, but the budget is an instructive research tool to pinpoint the major and minor pathways and transformations of N flow in mangrove ecosystems, and will help to identify where further research is needed.

The budget was constructed based on several assumptions: (1) the current (2014) global mangrove area is 86,495 km^2 [242]; (2) although mangrove tidal creeks and waterways are only a small fraction of total mangrove area, the total global mangrove area was used to calculate all tidal exchanges because the data includes many measurements taken from mangrove-fringed estuaries that are likely in toto to be of equivalent area; (3) litter export (PON) was estimated by converting litter C export [243] to N assuming a litter C/N of 79.4 (Table 1); (4) wood, litter, root and benthic microalgal NPP were estimated by converting the C values in [243] using the C/N ratios (g/g) in the Table 1 legend and using a microalgal C/N ratio (g/g) of 12 [244]; (5) standing stocks of soil and belowground roots and aboveground forest were derived from Figure 2; (6) the input of the total belowground root + soil pool (441,150 Gg N) to N burial (1127 Gg a^{-1}) was estimated by the difference between the mean N burial value (1239 Gg a^{-1}) minus the inputs from root production (62 Gg a^{-1}) and litter production (50 Gg a^{-1}); (7) wet N deposition was estimated from the only two studies available [4,245]; and (8) the differences between gross and net ammonification and nitrification represent N immobilization.

Not included in the budget are: (1) direct inputs from groundwater and upstream; (2) marine and terrigenous particle flux and deposition at the soil surface; (3) pelagic and benthic production; (4) dry deposition; (5) consumption and assimilation by fauna and flora; (6) N_2 -fixation on tree stems, cyanobacterial mats, aboveground roots, senescent leaves and litter; the latter are not included because of the inability to extrapolate these rates to more than a small area given the lack of knowledge of their areal coverage in a "typical" mangrove forest, and (7) rates of soil N transformations such as ammonification and denitrification likely account for only a part of total N flux in soils as most studies measured these rates only to soil depths of 5–20 cm. C mineralization is active to a soil depth of 1 m [243], so it is likely that significant N mineralization occurs in deeper soils.

Further, mangroves differ in their location and climate, and N-cycle pathways are significantly affected by physicochemical and biological factors, such as temperature, grazing, presence of biogenic structures, tides, pH, soil nutrients, soil type and rate of N input. Such variations have not, of course, been included in the mass balance, but must be kept in mind when considering the N cycle in mangroves. Thus, individual mangrove forests and ecosystems vary significantly from the global averages used here. For example, N cycling in a tropical mangrove ecosystem that is mature and luxuriant in terms of biomass and productivity is almost certainly to be more rapid and complex than N transformations in a subtropical fringing forest that is younger, less productive and smaller in terms of biomass.

Despite these significant shortcomings, the budget does suggest rapid rates of N cycling and transformations in mangrove forests, especially in the soil. Burial equates to about 29% of total N inputs. Net tidal exchange of dissolved N is into the forest or near zero, indicating N conservation. Anammox is as important a transformation process as denitrification. Denitrification equates to 35% of total N input, within the range found in other coastal ecosystems [246]. Net immobilization (8825 Gg N a⁻¹) is the single largest transformation process, underscoring its significance in conserving N in mangrove soils. In terms of N, benthic microalgae appear to be the most important primary producers, although the cyanobacterial mats are not represented. Net uptake of dissolved N by soils from overlying tidal waters is a significant N conservation mechanism, likely reflecting uptake by algae, cyanobacteria and other autotrophs on the soil surface. Alongi [11] measured high rates of dissolved N uptake by stems, logs, prop roots, and twigs, supporting the idea that microbial communities on these surfaces also function as an N conservation mechanism.

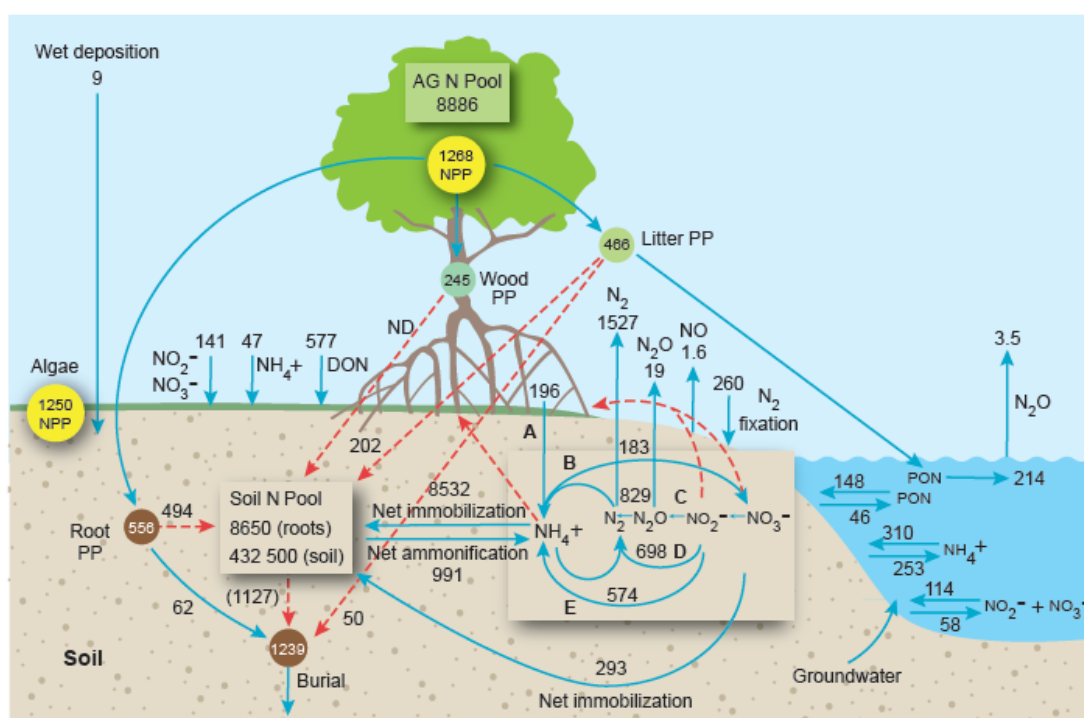


Figure 4. Nitrogen cycling in the world's mangrove ecosystems. Mean fluxes = Gg N a⁻¹; mean standing stocks = Gg N. The model assumes a global mangrove area of 86,495 km² [242]. Soil N transformations are lettered as: (A) root + rhizome N₂-fixation; (B) net nitrification; (C) denitrification; (D) anammox; (E) dissimilatory nitrate reduction to ammonium. Dashed red arrows represent mean values estimated indirectly (by difference); solid blue arrows represent mean values based on empirical measurements (see text for explanation and references). The N pool (both roots and soil) in soils to a depth of 1 m is presented as a box on left in the forest floor. N transformation in soils to depths of 5–20 cm is presented as a box on the right in the forest floor. Unquantified inputs and outputs of dissolved nitrogen from land-derived groundwater and organic matter inputs from adjacent marine waters and catchments are not represented. Abbreviations: ND = no data; AG N Pool = aboveground forest N biomass pool; PP = primary production; NPP = net primary production; NO₂⁻ + NO₃⁻ = nitrite plus nitrate; NH₄⁺ = ammonium; N₂ = gaseous nitrogen; N₂O = nitrous oxide; NO = nitric oxide; PON = particulate organic nitrogen.

The budget indicates a burial efficiency (burial/total input × 100) of 29% and a mineralization efficiency (gross ammonification-net ammonification/total input × 100) of 141%, which will be more realistically lower when other inputs (e.g., N₂-fixation from stems, etc.) are eventually included. In any case, it appears that soil N is very efficiently mineralized, as found in individual forests in

Asia [9,57,58,136]. For instance, in Chinese mangroves (*Kandelia candel*), burial efficiencies ranged from 8 to 31% and mineralization efficiencies from 69 to 92% [9]. In Thai mangroves, N burial efficiencies ranged from 4 to 12% and N mineralization efficiencies from 68 to 88% [57] and in Malaysian forests [136] burial and mineralization efficiencies ranged from 10 to 29% and 67 to 81%, respectively.

A mass balance of all inputs and outputs (Table 6) indicates a net positive gain of 957.9 Gg N a⁻¹ which equals 12% of all inputs and outputs, well within the sum of systematic errors of the many measurements made of N cycling. Considering that several important processes are not included, it is possible that N flow in mangrove ecosystems may be in approximate balance. However, considering the high rates of N₂-fixation on tree stems, cyanobacterial mats, aboveground roots, senescent leaves and litter, it is likely that ecosystem N inputs would need to be revised upwards when further studies with a proper sampling design incorporating the great spatial variability of this process (coupled to an appropriate computer algorithm) would warrant their inclusion in the N budget. The budget would be unbalanced with a larger net positive gain, but consumption and assimilation of these unquantified N₂-fixers by mangrove-associated fauna would perhaps redress the imbalance. Many organisms such as gastropods and other benthic invertebrates commonly dwell on tree stems, cyanobacterial mats, prop roots, pneumatophores, leaves and litter and readily consume organic particles, micro- and macroalgae, bacteria, and detritus on these surfaces [1,2]. Further, as pointed out by Reis et al. [247], N cycling in mangroves is unbalanced when forests are subjected to additional nutrients from anthropogenic sources, such as effluents from aquaculture ponds and human and animal sewage, so only relatively pristine mangroves would be expected to exhibit balanced N flow.

Table 6. A nitrogen mass balance of the world's mangrove ecosystems. Units = Gg N a⁻¹. Values are from Figure 4.

Inputs		Outputs	
Component	Gg N a ⁻¹	Component	Gg N a ⁻¹
Net primary production	1250	Burial	1239
Wood	245	Soil NO release	1.6
Litter	466	Soil N ₂ O release	19
Roots	556	Denitrification/Anammox	1527
Microalgae	1250	Water-air exchange	
N ₂ -fixation		N ₂ O	3.5
Roots	196	Tidal exchange	
Soil	260	DON	58
Precipitation	9	NO ₂ ⁻ + NO ₃ ⁻	46
Tidal exchange		NH ₄ ⁺	253
DON	148	PON	214
NO ₂ ⁻ + NO ₃ ⁻	114		
NH ₄ ⁺	310		
Soil-Water exchange			
DON	577		
NO ₂ ⁻ + NO ₃ ⁻	141		
NH ₄ ⁺	47		
Total	4319	TOTAL	3361.1
Inputs – Outputs = 957.9 (Net Gain)			

At the global scale, mangroves contribute variable percentages of N to the coastal ocean. Mangrove PON export (2474 kg N km⁻² a⁻¹) equates to 95% of PON export (2612 kg N km⁻² a⁻¹) from the world's major tropical rivers. However, most rivers in low latitudes have not been measured for PON export [248] so the true contribution is probably considerably smaller. Mangrove PON export accounts for only 1.5% of the entire world's river discharge [249]. The contributions of mangrove N₂O emissions,

denitrification, and burial to the global coastal ocean [250] are modest (0.4%, 0.5–2.0% and 6%, respectively), but are disproportionate relative to their small area (0.31%) [251].

6. Conclusions

N cycling in mangrove forests and associated tidal waterways is complex, with rapid turnover of low concentrations of dissolved N but slow turnover of particulate N; 96% of the latter is stored in soils with a total N stock of global mangrove forests of 52.03 Mg N ha⁻¹ which is considerably greater than their tropical terrestrial counterparts (22.33 Mg N ha⁻¹). The largest sources of N are nearly equal amounts of mangrove net primary production and production of benthic microalgae, the latter being a major source of nutrition for most mangrove-associated fauna. These benthic autotrophs are likely responsible for the net uptake of dissolved N species, mostly in the form of DON. Tides exchange dissolved and particulate N, with net uptake of dissolved N by mangroves and net release of particulate N as litter. N₂-fixation is an underestimated source of N; very high rates of N₂-fixation have been measured on tree stems, aboveground roots, and cyanobacterial mats but due to their very high patchiness and the lack of data to extrapolate beyond a small area, their contribution is unquantifiable at the whole-forest scale. Net immobilization, estimated as the difference between gross and net ammonification and nitrification, is the single largest (8825 Gg N a⁻¹) flux, reflecting its significance in conserving N. Denitrification is the largest loss pathway, equating to 35% of total N input, which is within the range measured in other coastal ecosystems. Burial in soil equates to about 29% of total N inputs and is the second largest loss of N. Overall, total inputs (4319 Gg N a⁻¹) slightly exceed total outputs (3361 Gg N a⁻¹) at present, suggesting net balance of N flow in mangrove ecosystems. Globally, mangrove PON export (2474 kg N km⁻² a⁻¹) equates to 95% of PON export (2612 kg N km⁻² a⁻¹) by the world's major tropical rivers; mangrove PON export accounts for only 1.5% of the entire world's river discharge. The contributions of mangrove N₂O emissions, denitrification, and burial to the global coastal ocean are modest (0.4%, 0.5–2.0% and 6%, respectively), but are disproportionate relative to their small area.

Despite these findings, many aspects of nitrogen flow in mangroves need to be properly quantified. These include: (1) the need to develop formulae to extrapolate N₂-fixation rates on stems, aboveground roots and cyanobacterial mats to the whole-forest scale; (2) measurement of N transformation processes in soils deeper than 20 cm; (3) direct groundwater inputs and inputs from upstream and marine sources, including deposition at the soil surface; (4) the contribution of fauna in N cycling, especially benthic crabs, epiphytes and plankton; and (5) wet and dry deposition.

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References

1. Alongi, D.M. *The Energetics of Mangrove Forests*; Springer Science: Dordrecht, The Netherlands, 2009.
2. Feller, I.C.; Lovelock, C.E.; Berger, U.; McKee, K.L.; Joye, S.B.; Ball, M.C. Biocomplexity in mangrove ecosystems. *Annu. Rev. Mar. Sci.* **2010**, *2*, 395–417. [[CrossRef](#)] [[PubMed](#)]
3. Alongi, D.M. Cycling and global fluxes of nitrogen in mangroves. *Glob. Environ. Res.* **2013**, *17*, 173–182.
4. Alongi, D.M.; Boto, K.G.; Robertson, A.I. Nitrogen and Phosphorus Cycles. In *Tropical Mangrove Ecosystems*; Robertson, A.I., Alongi, D.M., Eds.; American Geophysical Union: Washington, DC, USA, 1992; pp. 251–292.
5. Reef, R.; Feller, I.C.; Lovelock, C.E. Nutrition of mangroves. *Tree Physiol.* **2010**, *30*, 1148–1160. [[CrossRef](#)]
6. Boto, K.G. Nutrients and mangroves. In *Pollution in Tropical Aquatic Systems*; Connell, D.M., Hawker, D.W., Eds.; CRC Press: Boca Raton, FL, USA, 1991; pp. 129–145.
7. Lovelock, C.E.; Feller, I.C.; Ball, M.C.; Engelbrecht, B.M.J.; Ewe, M.L. Differences in plant function in phosphorus- and nitrogen-limited mangrove ecosystems. *New Phytol.* **2006**, *172*, 514–522. [[CrossRef](#)] [[PubMed](#)]

8. Watanabe, A.; Tsutsuki, K.; Inoue, Y.; Maie, N.; Melling, L.; Jaffé, R. Composition of dissolved organic nitrogen in rivers associated with wetlands. *Sci. Total Environ.* **2014**, *493*, 220–228. [[CrossRef](#)] [[PubMed](#)]
9. Alongi, D.M.; Pfitzner, J.; Trott, L.A.; Tirendi, F.; Dixon, P.; Klumpp, D.W. Rapid sediment accumulation and microbial mineralization in forests of the mangrove *Kandelia candel* in the Jiulongjiang Estuary, China. *Estuar. Coast. Shelf Sci.* **2005**, *63*, 605–618. [[CrossRef](#)]
10. Boto, K.G.; Wellington, J.T. Soil characteristics and nutrient status in northern Australian mangrove forests. *Estuaries* **1984**, *7*, 61–69. [[CrossRef](#)]
11. Alongi, D.M. The dynamics of benthic nutrient pools and fluxes in tropical mangrove forests. *J. Mar. Res.* **1996**, *54*, 123–148. [[CrossRef](#)]
12. Naidoo, G. Effects of salinity and nitrogen on growth and water relations in the mangrove, *Avicennia marina* (Forsk.) Vierh. *New Phytol.* **1987**, *107*, 317–325. [[CrossRef](#)]
13. Naidoo, G. Effects of nitrate, ammonium and salinity on growth of the mangrove *Bruguiera gymnorrhiza* (L.) Lam. *Aquat. Bot.* **1990**, *8*, 209–219. [[CrossRef](#)]
14. Shiau, Y.J.; Lee, S.C.; Chen, T.H.; Tian, G.; Chiu, C.Y. Water salinity effects on growth and nitrogen assimilation rate of mangrove (*Kandelia candel*) seedlings. *Aquat. Bot.* **2017**, *137*, 50–55. [[CrossRef](#)]
15. Zhao, Y.; Wang, X.; Wang, Y.; Jiang, Z.; Ma, X.; Inyang, A.I.; Cheng, H. Effects of salt on root aeration, nitrification, and nitrogen uptake in mangroves. *Forests* **2019**, *10*, 1131. [[CrossRef](#)]
16. Lalnunzira, C.; Tripathi, S.K. Leaf and root production, decomposition and carbon and nitrogen fluxes during stand development in tropical moist forests, north-east India. *Soil Res.* **2018**, *56*, 306–317. [[CrossRef](#)]
17. Leuschner, C.; Harteveld, M.; Hertel, D. Consequences of increasing forest use intensity for biomass, morphology, and growth of fine roots in a tropical moist forest on Sulawesi, Indonesia. *Agric. Ecosyst. Environ.* **2009**, *129*, 474–481. [[CrossRef](#)]
18. Valverde-Barrantes, O.J.; Raich, J.W.; Russell, A.E. Fine-root mass, growth and nitrogen content for six tropical tree species. *Plant Soil* **2007**, *290*, 357–370. [[CrossRef](#)]
19. Asante, W.; Jengre, N. Carbon Stocks and Soil Nutrient Dynamics in the Peat Swamp Forests of the Amanzule Wetlands and Ankobra River Basin. In *USAID Integrated Coastal and Fisheries Governance Program for the Western Region of Ghana*; Nature Conservation and Research Centre: Accra, Ghana, 2012; p. 45.
20. Vitousek, P.M.; Sanford, R.L., Jr. Nutrient cycling in moist tropical forest. *Annu. Rev. Ecol. Syst.* **1986**, *17*, 137–167. [[CrossRef](#)]
21. Martin, A.R.; Erickson, D.L.; Kress, W.J.; Thomas, S.C. Wood nitrogen concentrations in tropical trees: Phylogenetic patterns and ecological correlates. *New Phytol.* **2014**, *204*, 484–495. [[CrossRef](#)]
22. Herbohn, J.L.; Congdon, R.A. Ecosystem dynamics at disturbed and undisturbed sites in North Queensland wet tropical rain forest. III. Nutrient returns to the forest floor through litterfall. *J. Trop. Ecol.* **1998**, *14*, 217–229. [[CrossRef](#)]
23. Msunaga, T.; Kubota, D.; Hotta, M.; Wakatsuki, T. Nutritional characteristics of mineral elements in tree species of tropical rain forest, West Sumatra, Indonesia. *Soil Sci. Plant Nutr.* **1997**, *43*, 405–418. [[CrossRef](#)]
24. Paoli, G.D.; Curran, L.M. Soil nutrients limit fine litter production and tree growth in mature lowland forest of southwestern Borneo. *Ecosystems* **2007**, *10*, 503–518. [[CrossRef](#)]
25. Sugihara, S.; Shibata, M.; Mvondo Ze, A.D.; Araki, S.; Funakawa, S. Effect of vegetation on soil C, N, P and other minerals in oxisols at the forest-savanna transition zone on central Africa. *Soil Sci. Plant Nutr.* **2014**, *60*, 45–59. [[CrossRef](#)]
26. Martinelli, L.A.; Piccolo, M.C.; Townsend, A.R.; Vitousek, P.M.; Cuevas, E.; McDowell, W.; Robertson, G.P.; Santos, O.C.; Treseder, K. Nitrogen stable isotopic composition of leaves and soil: Tropical versus temperate forests. *Biogeochemistry* **1999**, *46*, 45–65. [[CrossRef](#)]
27. Schurr, E.A.G.; Matson, P.A. Net primary productivity and nutrient cycling across a mesic to wet precipitation gradient in Hawaiian montane forest. *Oecologia* **2001**, *128*, 431–442. [[CrossRef](#)] [[PubMed](#)]
28. Proctor, J.; Anderson, J.; Chai, P.; Vallack, H.W. Ecological studies in four contrasting lowland rain forests in Gunung Mulu National Park, Sarawak. I. Forest environment, structure and floristics. *J. Ecol.* **1983**, *71*, 237–260. [[CrossRef](#)]
29. Shibata, M.; Sugihara, S.; Mvondo-Ze, A.D.; Taraki, S.; Funakawa, S. Nitrogen flux patterns through oxisols and ultisols in tropical forests of Cameroon, central Africa. *Soil Sci. Plant Nutr.* **2017**, *63*, 306–317. [[CrossRef](#)]
30. Weintraub, S.R.; Taylor, P.G.; Porder, S.; Asner, G.P. Topographic controls on soil nitrogen availability in a lowland tropical forest. *Ecology* **2015**, *96*, 1561–1574. [[CrossRef](#)]

31. Reed, S.C.; Cleveland, C.C.; Townsend, A.R. Tree species control rates of free-living nitrogen fixation in a tropical rain forest. *Ecology* **2008**, *89*, 2924–2934. [[CrossRef](#)]
32. Tuah, S.J.; Jamal, Y.M.; Limin, S.H. Nutritional characteristics in leaves of plants native to tropical peat swamps and heath forests of central Kalimantan, Indonesia. *Tropics* **2003**, *12*, 224–232. [[CrossRef](#)]
33. Powers, J.S.; Schlesinger, W.H. Relationships among soil carbon distributions and biophysical factors at nested spatial scales in rain forests of northeastern Costa Rica. *Geoderma* **2002**, *109*, 165–190. [[CrossRef](#)]
34. Werner, C.; Kiese, R.; Butterbach-Bahl, K. Soil-atmosphere exchange of N₂O, CH₄, and CO₂ and controlling environmental factors for tropical rain forest sites in western Kenya. *J. Geophys. Res.* **2007**, *112*, D03308. [[CrossRef](#)]
35. Koehler, B.; Corre, M.D.; Veldkamp, E.; Wullaert, H.; Wright, S.J. Immediate and long-term nitrogen oxide emissions from tropical forest soils exposed to elevated nitrogen input. *Glob. Chang. Biol.* **2009**, *15*, 2049–2066. [[CrossRef](#)]
36. Neto, E.S.; Carmo, J.B.; Keller, M.; Martins, S.C.; Alves, L.F.; Vieira, S.A.; Piccolo, M.C.; Camargo, P.; Couto, H.T.Z.; Joly, C.A.; et al. Soil-atmosphere exchange of nitrous oxide, methane and carbon dioxide in a gradient of elevation in the coastal Brazilian Atlantic forest. *Biogeosciences* **2011**, *8*, 733–742. [[CrossRef](#)]
37. Palmiotto, P.A.; Davies, S.J.; Vogt, K.A.; Ashton, M.S.; Vogt, D.J.; Ashton, P.S. Soil-related habitat specialization in dipterocarp rain forest tree species in Borneo. *J. Ecol.* **2004**, *92*, 609–623. [[CrossRef](#)]
38. Vance, E.D.; Nadkarni, N.M. Microbial biomass and activity in canopy organic matter and the forest floor of a tropical cloud forest. *Soil Biol. Biochem.* **1990**, *22*, 677–684. [[CrossRef](#)]
39. Alongi, D.M. Global significance of mangrove blue carbon in climate change mitigation. *Sci* **2020**, *2*, 67. [[CrossRef](#)]
40. Naidoo, G. Differential effects of nitrogen and phosphorus enrichment on growth of dwarf *Avicennia marina* mangroves. *Aquat. Bot.* **2009**, *90*, 184–190. [[CrossRef](#)]
41. Lovelock, C.E.; Ruess, R.W.; Feller, I.C. Fine root respiration in the mangrove *Rhizophora mangle* over variation in forest stature and nutrient availability. *Tree Physiol.* **2006**, *26*, 1601–1606. [[CrossRef](#)]
42. Lee, R.Y.; Porubsky, W.P.; Feller, I.C.; Mc Kee, K.L.; Joye, S.B. Porewater biogeochemistry and soil metabolism in dwarf red mangrove habitats (Twin Cays, Belize). *Biogeochemistry* **2008**, *87*, 181–198. [[CrossRef](#)]
43. Lacerda, L.D.; Ittekkot, V.; Patchineelam, S.R. Biogeochemistry of mangrove soil organic matter: A comparison between *Rhizophora* and *Avicennia* soils in south-eastern Brazil. *Estuar. Coast. Shelf Sci.* **1995**, *40*, 713–720. [[CrossRef](#)]
44. Chen, R.; Twilley, R.R. A simulation model of organic matter and nutrient accumulation in mangrove wetland soils. *Biogeochemistry* **1999**, *44*, 93–118. [[CrossRef](#)]
45. Krauss, K.W.; Doyle, T.W.; Twilley, R.R.; Rivera-Monroy, V.H.; Sullivan, J.K. Evaluating the relative contributions of hydroperiod and soil fertility on growth of south Florida mangroves. *Hydrobiologia* **2006**, *569*, 311–324. [[CrossRef](#)]
46. Rovai, A.S.; Twilley, R.R.; Castañeda-Moya, E.; Riul, P.; Cifuentes-Jara, M.; Manrow-Villalobos, M.; Horta, P.A.; Simonassi, J.C.; Fonseca, A.L.; Pagliosa, P.R. Global controls on carbon storage in mangrove soils. *Nat. Clim. Chan.* **2018**, *3*, 534–538. [[CrossRef](#)]
47. Bala Krishna Prasad, M.; Ramanathan, A.L. Sedimentary nutrient dynamics in a tropical estuarine mangrove ecosystem. *Estuar. Coast. Shelf Sci.* **2008**, *80*, 60–66. [[CrossRef](#)]
48. Alongi, D.M.; Tirendi, F.; Clough, B.F. Below-ground decomposition of organic matter in forests of the mangroves *Rhizophora stylosa* and *Avicennia marina* along the arid coast of Western Australia. *Aquat. Bot.* **2000**, *68*, 97–122. [[CrossRef](#)]
49. Alongi, D.M.; Christoffersen, P.; Tirendi, F. The influence of microbial-nutrient relationships in tropical mangrove sediments. *J. Exp. Mar. Biol. Ecol.* **1993**, *171*, 201–223. [[CrossRef](#)]
50. Alongi, D.M.; de Carvalho, N.A. The effect of small-scale logging on stand characteristics and soil biogeochemistry in mangrove forests of Timor Leste. *For. Ecol. Manag.* **2008**, *255*, 1359–1366. [[CrossRef](#)]
51. Alongi, D.M.; Tirendi, F.; Dixon, P.; Trott, L.A.; Brunskill, G.J. Mineralization of organic matter in intertidal sediments of a tropical semi-enclosed delta. *Estuar. Coast. Shelf Sci.* **1999**, *48*, 451–467. [[CrossRef](#)]
52. Alongi, D.M.; Ramanathan, A.L.; Kannan, L.; Tirendi, F.; Trott, L.A.; Bala Krishna Prasad, M. Influence of human-induced disturbance on benthic microbial metabolism in the Pichavaram mangroves, Vellar-Coleroon estuarine complex, India. *Mar. Biol.* **2005**, *147*, 1033–1044. [[CrossRef](#)]

53. Alongi, D.M. Bacterial productivity and microbial biomass in tropical mangrove sediments. *Microb. Ecol.* **1988**, *15*, 59–79. [[CrossRef](#)]
54. Alongi, D.M.; de Carvalho, N.A.; Amaral, A.L.; da Costa, A.; Trott, L.A.; Tirendi, F. Uncoupled surface and below-ground soil respiration in mangroves: Implications for estimates of dissolved inorganic carbon export. *Biogeochemistry* **2012**, *109*, 151–162. [[CrossRef](#)]
55. Alongi, D.M.; Trott, L.A.; Tirendi, F.; McKinnon, A.D.; Undu, M.C. Growth and development of mangrove forests overlying smothered coral reefs, Sulawesi and Sumatra, Indonesia. *Mar. Ecol. Prog. Ser.* **2008**, *370*, 97–109. [[CrossRef](#)]
56. Alongi, D.M.; Tirendi, F.; Trott, L.A.; Xuan, T.T. Benthic decomposition rates and pathways in plantations of the mangrove *Rhizophora apiculata* in the Mekong delta, Vietnam. *Mar. Ecol. Prog. Ser.* **2000**, *194*, 87–101. [[CrossRef](#)]
57. Alongi, D.M.; Trott, L.A.; Wattayakorn, G.; Clough, B.F. Below-ground nitrogen cycling in relation to net canopy production in mangrove forests of southern Thailand. *Mar. Biol.* **2002**, *140*, 855–864.
58. Kristensen, E.; Holmer, M.; Banta, G.T.; Jensen, M.H.; Hansen, K. Carbon, nitrogen and sulfur cycling in sediments of the Ao Nam Bor mangrove forest, Phuket, Thailand: A review. *Phuket Mar. Biol. Cent. Res. Bull.* **1995**, *60*, 37–64.
59. Nguyen, H.T.; Yoneda, R.; Ninomiya, I.; Harada, K.; Van Dao, T.; Sy, T.M.; Phan, H.N. The effects of stand-age and inundation on carbon accumulation in mangrove plantation soil in Namdinh, northern Vietnam. *Tropics* **2004**, *14*, 24–34. [[CrossRef](#)]
60. Castañeda-Moya, E.; Twilley, R.R.; Rivera-Monroy, V.H.; Zhang, K.; Davis, S.E., III; Ross, M. Sediment and nutrient deposition associated with Hurricane Wilma in mangroves of the Florida Coastal Everglades. *Estuar. Coast.* **2010**, *33*, 45–58. [[CrossRef](#)]
61. Garcias-Bonet, N.; Delgado-Huertas, A.; Carrillo-de-Albornoz, P.; Anton, A.; Almahasheer, H.; Marbá, N.; Hendricks, I.E.; Krause-Jensen, D.; Duarte, C.M. Carbon and nitrogen concentrations, stocks, and isotopic compositions in Red Sea seagrass and mangrove sediments. *Front. Mar. Sci.* **2019**, *6*, 267. [[CrossRef](#)]
62. Rivera-Monroy, V.H.; Twilley, R.R.; Medina, E.; Moser, E.B.; Botero, L.; Francisco, A.M.; Bullard, E. Spatial variability of soil nutrients in disturbed riverine mangrove forests at different stages of regeneration in the San Juan River estuary, Venezuela. *Estuaries* **2004**, *27*, 44–57. [[CrossRef](#)]
63. Gandaseca, S.; Pazi, A.M.M.; Zulkipli, M.N.S.; Hamzah, A.H.; Zaki, P.H.; Abdu, A. Assessment of nitrogen and phosphorus in mangrove forest soil at Awat-Awat Lawas Sarawak. *Am. J. Agricult. For.* **2016**, *4*, 136–139. [[CrossRef](#)]
64. Sarker, S.; Masud-Ul-Alam, M.; Hossain, M.S.; Chowdhury, S.R.; Sharifuzzaman, S.M. A review of bioturbation and sediment organic geochemistry in mangroves. *Geol. J.* **2020**. [[CrossRef](#)]
65. Sofawi, A.B.; Nazri, M.N.; Rozainah, M.Z. Nutrient availability in mangrove soil: Anthropogenic, seasonal and depth variation factors. *Appl. Ecol. Environ. Res.* **2017**, *15*, 1983–1998. [[CrossRef](#)]
66. Shilla, D.J.; Shilla, D.A. Assessment of the geochemical characteristics of water and surface sediments of Rufiji mangrove forest, Tanzania. *Tanzan. J. Sci.* **2020**, *46*, 482–497.
67. Gutiérrez, J.C.S.; Ponce-Palafox, J.T.; Pineda-Jaimes, N.B.; Arenas-Fuentes, V.; Arredondo-Figueroa, J.L.; Cifuentes-Lemus, J.L. Comparison of the mangrove soil with different levels of disturbance in tropical Agua Brava Lagoon, Mexican Pacific. *Appl. Ecol. Environ. Res.* **2016**, *14*, 45–57. [[CrossRef](#)]
68. Ramanathan, A.L.; Datta, D.K.; Ghosh, P.; Kaushal, S.; Murtudde, R. *Tracing Nitrogen and Carbon Biogeochemical Processes in the Intertidal Mangrove Ecosystem (Sundarbans) of India and Bangladesh: Implications for the Global Environmental Change*; Final Report for APN Project: ARCP2012-07CMY-Ramanathan; Asia-Pacific Network for Global Change Research: Kobe, Japan, 2012.
69. Scharler, U.M.; Ulanowicz, R.E.; Fogel, M.L.; Wooller, M.J.; Jacobson-Meyers, M.E.; Lovelock, C.E.; Feller, I.C.; Frischer, M.; Lee, R.; McKee, K.; et al. Variable nutrient stoichiometry (carbon:nitrogen:phosphorus) across trophic levels determines community and ecosystem properties in an oligotrophic mangrove system. *Oecologia* **2015**, *179*, 863–876. [[CrossRef](#)]
70. Nirmal Kumar, I.J.; Sajish, P.R.; Nirmal Kumar, R.; Bash, G.; Shailendra, V. Nutrient dynamics in an *Avicennia marina* (Forsk.) Vierh., mangrove forest in Vamleshwar, Gujarat, India. *Not. Sci. Biol.* **2011**, *3*, 51–56. [[CrossRef](#)]
71. Nordhaus, I.; Salewski, T.; Jennerjahn, T.C. Interspecific variations in mangrove leaf litter decomposition are related to labile nitrogenous compounds. *Estuar. Coast. Shelf Sci.* **2017**, *192*, 137–143. [[CrossRef](#)]

72. Bulmer, R.H.; Schwendenmann, L.; Lundquist, C.J. Carbon and nitrogen stocks and below-ground allometry in temperate mangroves. *Front. Mar. Sci.* **2016**, *3*, 150. [[CrossRef](#)]
73. Alongi, D.M.; Clough, B.F.; Dixon, P.; Tirendi, F. Nutrient partitioning and storage in arid-zone forests of the mangroves *Rhizophora stylosa* and *Avicennia marina*. *Trees* **2003**, *17*, 51–60. [[CrossRef](#)]
74. Marchand, C.; Lallier-Vergès, E.; Baltzer, F. The composition of sedimentary organic matter in relation to the dynamic features of the mangrove-fringed coast of French Guiana. *Estuar. Coast. Shelf Sci.* **2003**, *56*, 119–130. [[CrossRef](#)]
75. Enamul Hoq, M.; Islam, M.L.; Paul, H.K.; Ahmed, S.U.; Islam, M.N. Decomposition and seasonal changes in nutrient constituents in mangrove litter of Sundarbans mangrove, Bangladesh. *Ind. J. Mar. Sci.* **2002**, *31*, 130–135.
76. Haryadi, J.; Basukriandi, A. The study on mangrove litter as a source of nutrients for Blanakan mangrove pond, Subang, West Java. *Indo. Aquacult. J.* **2013**, *8*, 55–64.
77. Nordhaus, I.; Salewski, T.; Jennerjahn, T.C. Food preferences on mangrove crabs related to leaf nitrogen compounds in the Segara Anakan Lagoon, Java, Indonesia. *J. Sea Res.* **2011**, *65*, 414–426. [[CrossRef](#)]
78. Twilley, R.R.; Lugo, A.E.; Patterson-Zucca, C. Litter production and turnover in basin mangrove forests in southwest Florida. *Ecology* **1986**, *67*, 670–683. [[CrossRef](#)]
79. Wafar, S.; Untawale, A.G.; Wafar, M. Litter fall and energy flux in a mangrove ecosystem. *Estuar. Coast. Shelf Sci.* **1997**, *44*, 111–124. [[CrossRef](#)]
80. Hemati, Z.; Hossian, M.; Rozainah, M.Z. Determination of carbon and nitrogen in litter fall of mangrove ecosystem in Peninsular Malaysia. *Pak. J. Bot.* **2017**, *49*, 1381–1386.
81. Ye, Y.; Chen, Y.P.; Chen, G.C. Litter production and litter elemental composition in two rehabilitated *Kandelia obovata* mangrove forests in Jiulongjiang estuary, China. *Mar. Environ. Res.* **2013**, *83*, 63–72. [[CrossRef](#)]
82. Srisunont, C.; Jaiyen, T.; Tenrung, M.; Likitchaikul, M.; Srisunont, T. Nutrient accumulation by litterfall in mangrove forest at Klong Khone, Thailand. *Sci. Technol. Asia* **2017**, *9*–18. [[CrossRef](#)]
83. Adame, M.F.; Zacaria, R.M.; Fry, B.; Chong, V.C.; Then, Y.H.A.; Brown, C.J.; Lee, S.Y. Loss and recovery of carbon and nitrogen after mangrove clearing. *Ocean. Coast. Manag.* **2018**, *161*, 117–126. [[CrossRef](#)]
84. Lugo, A.E.; Medina, E.; Cuevas, E.; Laboy Nieves, E.N.; Schaeffer Novelli, Y. Ecophysiology of a mangrove forest in Jobos Bay, Puerto Rico. *Carrib. J. Sci.* **2007**, *43*, 200–219. [[CrossRef](#)]
85. Pinto, L. Litterfall and its element content in the Pagbilao Mangrove Forest Reserve, Philippines. *Mahasagar* **1992**, *25*, 97–104.
86. Pedrosa Fragoso, C.; Bernini, E.; Ferreira Araújo, B.; Gomes de Almeida, M.; Eduardo de Rezende, C. Mercury in litterfall and sediment using elemental and isotopic composition of carbon and nitrogen in the mangrove of southeastern Brazil. *Estuar. Coast. Shelf Sci.* **2018**, *202*, 30–39. [[CrossRef](#)]
87. Bunt, J.S. Studies of Mangrove Litter Fall in Tropical Australia. In *Mangrove Ecosystems in Australia: Structure, Function and Management*; Clough, B.F., Ed.; Australian National University Press: Canberra, Australia, 1982; pp. 221–237.
88. Woodroffe, C.D.; Bardsley, K.N.; Ward, P.J.; Hanley, J.R. Production of mangrove litter in a macrotidal embayment, Darwin Harbour, N.T., Australia. *Estuar. Coast. Shelf Sci.* **1988**, *26*, 581–598. [[CrossRef](#)]
89. Thongtham, N.; Kristensen, E. Carbon and nitrogen balance of leaf-eating sesarmid crabs (*Neopisesarma versicolor*) offered different food sources. *Estuar. Coast. Shelf Sci.* **2005**, *65*, 213–222. [[CrossRef](#)]
90. Thongtham, N.; Kristensen, E.; Puangprasan, S.-Y. Leaf removal by sesarmid crabs in Bangrong mangrove forests, Phuket, Thailand; with emphasis on the feeding ecology of *Neopisesarma versicolor*. *Estuar. Coast. Shelf Sci.* **2008**, *80*, 583–590. [[CrossRef](#)]
91. Feller, I.C.; McKee, K.L.; Whigham, D.F.; O'Neill, J.P. Nitrogen vs. phosphorus limitation across an ecotonal gradient in a mangrove forest. *Biogeochemistry* **2002**, *62*, 145–175. [[CrossRef](#)]
92. Saravanakumar, A.; Rajkumar, M.; Sesh Serebiah, J.; Thivakaran, G.A. Seasonal variations in physico-chemical characteristics of water, sediment and soil texture in arid zone mangroves of Kachchh-Gujarat. *J. Environ. Biol.* **2008**, *29*, 725–732.
93. Nabiul Islam Khan, M.; Suwa, R.; Hagihara, A. Carbon and nitrogen pools in a mangrove stand of *Kandelia obovata* (S.L.) Yong: Vertical distribution in the soil-vegetation system. *Wetlands Ecol. Manag.* **2007**, *15*, 141–153. [[CrossRef](#)]
94. Mantiquilla, J.A.; Salmasan, S.F.D.; Obelidhon, M.K.A.; Abad, R.G. Nutrient status of Nipa (*Nypa fruticans* Wurm.) in selected areas of Mindanao, the Philippines. *Banwa B* **2019**, *14*, art012.

95. Medina, E.; Francisco, M. Osmolality and $\delta^{13}\text{C}$ of leaf tissues of mangrove species from environments of contrasting rainfall and salinity. *Estuar. Coast. Shelf Sci.* **1997**, *45*, 337–344. [[CrossRef](#)]
96. Alongi, D.M.; Wattayakorn, G.; Tirendi, F.; Dixon, P. Nutrient capital in different aged forests of the mangrove *Rhizophora apiculata*. *Bot. Mar.* **2004**, *47*, 116–124. [[CrossRef](#)]
97. Tognella, M.M.P.; Soares, M.L.G.; Cuevas, E.; Medina, E. Heterogeneity of elemental composition and natural abundance of stable isotopes of C and N in soils and leaves of mangroves at their southernmost West Atlantic range. *Braz. J. Biol.* **2016**, *76*, 994–1003. [[CrossRef](#)]
98. Jayasekera, R. Chemical composition of the mangrove, *Rhizophora mangle* L. *J. Plant Physiol.* **1991**, *138*, 119–121. [[CrossRef](#)]
99. Feller, I.C.; Lovelock, C.E.; McKee, K.L. Nutrient addition differentially affects ecological processes of *Avicennia germinans* in nitrogen versus phosphorus limited mangrove ecosystems. *Ecosystems* **2007**, *10*, 347–359. [[CrossRef](#)]
100. Telave, A.B. Ecophysiological studies on *Sonneratia* L. from the coast of Maharashtra, India. *Ind. J. Geo-Mar. Sci.* **2015**, *44*, 1239–1244.
101. Muzuka, A.N.N.; Shunula, J.P. Stable isotope compositions of organic carbon and nitrogen of two mangrove stands along the Tanzanian coastal zone. *Estuar. Coast. Shelf Sci.* **2006**, *66*, 447–458. [[CrossRef](#)]
102. Bernini, E.; Silva, M.A.B.D.; Carmo, T.M.S.D.; Cuzzuol, G.R.F. Composição química do sedimento e de folhas das espécies do manguezal do estuário do Rio São Mateus, Espírito Santo, Brasil. *Rev. Brasil Bot.* **2006**, *29*, 689–699. [[CrossRef](#)]
103. Medina, E.; Giarrizzo, T.; Menezes, M.; Carvalho Lira, M.; Carvalho, E.A.; Peres, A.; Silva, B.; Vilhena, R.; Reise, A.A.; Braga, F.C. Mangal communities of the “Salgado Paraense”: Ecological heterogeneity along the Bragança peninsula assessed through soil and leaf analysis. *Amazonia* **2001**, *16*, 397–416.
104. Lovelock, C.E.; Feller, I.C.; McKee, K.L.; Engelbrecht, B.M.J.; Ball, M.C. The effect of nutrient enrichment on growth, photosynthesis and hydraulic conductance of dwarf mangroves in Panama. *Funct. Ecol.* **2004**, *18*, 25–33. [[CrossRef](#)]
105. Lin, Y.-M.; Liu, X.-W.; Zhang, H.; Fan, H.-Q.; Lin, G.-H. Nutrient conservation strategies of a mangrove species *Rhizophora stylosa* under nutrient limitation. *Plant Soil* **2010**, *326*, 469–479. [[CrossRef](#)]
106. Gritcan, I.; Duxbury, M.; Leuzinger, S.; Alfaro, A.C. Leaf stable isotope and nutrient status of temperate mangroves as ecological indicators to assess anthropogenic activity and recovery from eutrophication. *Front. Plant Sci.* **2016**, *7*, 1922. [[CrossRef](#)]
107. Simpson, L.T.; Lovelock, C.E.; Cherry, J.A.; Feller, I.C. Short-lived effects of nutrient enrichment on *Avicennia germinans* decomposition in a saltmarsh-mangrove ecotone. *Estuar. Coast. Shelf Sci.* **2020**, *235*, 106598. [[CrossRef](#)]
108. Ariyanto, D.; Gunawan, H.; Puspitasari, D.; Susanti Ningsih, S.; Jayanegara, A.; Hamim, H. The differences of the element content in *Rhizophora mucronata* leaves from Asahan Regency, North Sumatra, Indonesia. *Pol. J. Natur. Sci.* **2019**, *34*, 481–491.
109. Li, M.S. Nutrient dynamics of a Futian mangrove forest in Shenzhen, South China. *Estuar. Coast. Shelf Sci.* **1997**, *45*, 463–472. [[CrossRef](#)]
110. Bernini, E.; da Silva, M.A.B.; do Carmo, T.M.S.; Cuzzuol, G.R.F. Spatial and temporal variation of the nutrients in the sediment and leaves of two Brazilian mangrove species and their role in the retention of environmental heavy metals. *Braz. J. Plant Physiol.* **2010**, *22*, 177–187. [[CrossRef](#)]
111. Ahmed, A.; Ohlson, M.; Hoque, S.; Moula, M.G. Chemical composition of leaves of a mangrove tree (*Sonneratia apetala* Buch.Ham.) and their correlation with some soil variables. *Bangladesh J. Bot.* **2010**, *39*, 61–69. [[CrossRef](#)]
112. Mfilinge, P.L.; Atta, N.; Tsuchiya, M. Nutrient dynamics and leaf litter decomposition in a subtropical mangrove forest at Oura Bay, Okinawa, Japan. *Trees* **2002**, *16*, 172–180. [[CrossRef](#)]
113. Saatchi, S.S.; Harris, N.L.; Brown, S.; Lefsky, M.; Mitchard, E.T.A.; Salas, W.; Zutta, B.R.; Buermann, W.; Lewis, S.L.; Hagen, S.; et al. Benchmark map of forest carbon stocks in tropical regions across three continents. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 9899–9904. [[CrossRef](#)] [[PubMed](#)]
114. Sullivan, M.J.P.; Talbot, J.; Lewis, S.L.; Phillips, O.L.; Qie, L.; Begne, S.K.; Chave, J.; Cuni-Sanchez, A.; Hubau, W.; Lopez-Gonzalez, G.; et al. Diversity and carbon storage across the tropical forest biome. *Sci. Rep.* **2017**, *7*, 39102. [[CrossRef](#)]

115. Wantzen, K.M.; Couto, E.G.; Mund, E.E.; Amorim, R.S.S.; Siqueira, A.; Tielbörger, K.; Seifan, M. Soil carbon stocks in stream-valley-ecosystems in the Brazilian Cerrado agroscape. *Agric. Ecosyst. Environ.* **2012**, *151*, 70–79. [[CrossRef](#)]
116. Descloux, S.; Chanudet, V.; Poilvé, H.; Grégoire, A. Co-assessment of biomass and soil organic carbon stocks in a future reservoir area located in Southeast Asia. *Environ. Monit. Assess.* **2011**, *173*, 723–741. [[CrossRef](#)]
117. Laumonier, Y.; Edin, A.; Kanninen, M.; Munandar, A.W. Landscape-scale variation in the structure and biomass of the hill dipterocarp forest of Sumatra: Implications for carbon stock assessments. *For. Ecol. Manag.* **2010**, *259*, 505–513. [[CrossRef](#)]
118. Yuen, J.Q.; Ziegler, A.D.; Webb, E.L.; Ryan, C.M. Uncertainty in below-ground carbon biomass for major land covers in Southeast Asia. *For. Ecol. Manag.* **2013**, *310*, 915–926. [[CrossRef](#)]
119. Powers, J.S.; Treseder, K.K.; Lerdau, M.T. Fine roots, arbuscular mycorrhizal hyphae, and soil nutrients in four neotropical rain forests: Patterns across large geographic distances. *New Phytol.* **2005**, *165*, 913–921. [[CrossRef](#)]
120. Cusack, D.F.; Markesteijn, L.; Condit, R.; Lewis, O.T.; Turner, B.L. Soil carbon stocks across tropical forests of Panama regulated by base cation effects on fine roots. *Biogeochemistry* **2018**, *137*, 253–266. [[CrossRef](#)]
121. Thamdrup, B. New pathways and processes in the global nitrogen cycle. *Annu. Rev. Ecol. Evol. Syst.* **2012**, *43*, 407–428. [[CrossRef](#)]
122. Mukherji, S.; Haldar, S.; Ghosh, A. Investigation of the Structural and Functional Microbial Diversity in Indian Mangroves. In *Microorganisms in Saline Environments: Strategies and Functions*; Giri, B., Varma, A., Eds.; Springer Nature: Gland, Switzerland, 2019; pp. 93–130.
123. Cheung, M.K.; Wong, C.K.; Chu, K.H.; Kwan, H.S. Community structure, dynamics and interactions of bacteria, archaea and fungi in subtropical coastal wetland sediments. *Sci. Rep.* **2018**, *8*, 14397. [[CrossRef](#)] [[PubMed](#)]
124. Loganathachetti, D.S.; Poosakkannu, A.; Muthuraman, S. Fungal community assemblage of different soil compartments in mangrove ecosystem. *Sci. Rep.* **2017**, *7*, 8560. [[CrossRef](#)] [[PubMed](#)]
125. Marcos, M.S.; Barboza, A.D.; Keijzer, R.M.; Laanbroek, H.J. Tide as steering factor in structuring archaeal and bacterial ammonia-oxidizing communities in mangrove forest soils dominated by *Avicennia germinans* and *Rhizophora mangle*. *Microb. Ecol.* **2018**, *75*, 997–1008. [[CrossRef](#)]
126. Li, R.; Tong, T.; Wu, S.; Chai, M.; Xie, S. Multiple factors govern the biogeographic distribution of archaeal community in mangrove across China. *Estuar. Coast. Shelf Sci.* **2019**, *231*, 106414. [[CrossRef](#)]
127. Bai, S.; Li, J.; He, Z.; Van Nostrand, J.D.; Tian, Y.; Lin, G.; Zhou, J.; Zheng, T. GeoChip-based analysis of the functional gene diversity and metabolic potential of soil microbial communities in mangroves. *Appl. Microbiol. Biotechnol.* **2013**, *97*, 7035–7048. [[CrossRef](#)]
128. Zhang, M.; Luo, Y.; Lin, L.; Lin, X.; Hetharua, B.; Zhao, W.; Zhou, M.; Zhan, Q.; Xu, H.; Zheng, T.; et al. Molecular and stable isotope evidence for the occurrence of nitrite-dependent anaerobic methane-oxidizing bacteria in the mangrove sediment of Zhangjiang estuary, China. *Appl. Microbiol. Biotechnol.* **2018**, *102*, 2441–2454. [[CrossRef](#)] [[PubMed](#)]
129. Kristensen, E.; Jensen, M.H.; Banta, G.T.; Hansen, K.; Holmer, M.; King, G.M. Transformation and transport of inorganic nitrogen in sediments of a southeast Asian mangrove forest. *Aquat. Microb. Ecol.* **1998**, *15*, 165–175. [[CrossRef](#)]
130. Viner, A.B. The Status and Transport of Nutrients through the Purari River (Papua New Guinea). In *Purari River Hydroelectric Scheme Environmental Studies*; Publication of the Environment and Conservation: Waigani, Papua New Guinea, 1979; Volume 9, pp. 1–28.
131. Kyaruzi, J.J.; Kyewalyanga, M.S.; Muruke, M.H.S. Cyanobacteria composition and impact of seasonality on their *in situ* nitrogen fixation rate in a mangrove ecosystem adjacent to Zanzibar town. *West. Indian Ocean J. Mar. Sci.* **2003**, *2*, 35–44. [[CrossRef](#)]
132. van der Valk, A.G.; Attiwill, P.M. Acetylene reduction in an *Avicennia marina* community in southern Australia. *Aust. J. Bot.* **1984**, *32*, 157–164.
133. Lee, R.Y.; Joye, S.B. Seasonal patterns of nitrogen fixation and denitrification in oceanic mangrove habitats. *Mar. Ecol. Prog. Ser.* **2006**, *307*, 127–141. [[CrossRef](#)]
134. Joye, S.B.; Lee, R.Y. Benthic microbial mats: Important sources of fixed nitrogen and carbon to the Twin Cays, Belize ecosystem. *Atoll Res. Bull.* **2004**, *528*, 1–26. [[CrossRef](#)]

135. Whigham, D.F.; Verhoeven, J.T.A.; Samarkin, V.; Megonigal, P.J. Responses of *Avicennia germinans* (Black Mangrove) and the soil microbial community to nitrogen addition in a hypersaline wetland. *Estuar. Coast.* **2009**, *32*, 926–936. [[CrossRef](#)]
136. Alongi, D.M.; Sasekumar, A.; Chong, V.C.; Pfitzner, J.; Trott, L.A.; Tirendi, F.; Dixon, P.; Brunskill, G.J. Sediment accumulation and organic material flux in a managed mangrove ecosystem: Estimates of land-ocean-atmosphere exchange in peninsular Malaysia. *Mar. Geol.* **2004**, *208*, 383–402. [[CrossRef](#)]
137. Twilley, R.R.; Rivera-Monroy, V.H.; Rovai, A.S.; Castañeda-Moya, E.; Davis, S. Mangrove Biogeochemistry at Local and Global Scales Using Ecogeomorphic Approaches. In *Coastal Wetlands: An Integrated Ecosystem Approach*; Perillo, G.M.E., Wolanski, E., Cahoon, D.R., Hopkinson, C.S., Eds.; Elsevier: Amsterdam, The Netherlands, 2019; pp. 717–785.
138. Morell, J.M.; Corredor, J.E. Sediment trapping in a mangrove lagoon. *Estuar. Coast. Shelf Sci.* **1993**, *37*, 203–212. [[CrossRef](#)]
139. Das, S.; Ganguly, D.; Mukherjee, A.; Chakraborty, S.; De, T.K. Exploration of N₂ fixation and denitrification processes in the Sundarbans mangrove ecosystem, India. *Ind. J. Geo-Mar. Sci.* **2020**, *49*, 740–747.
140. Pelegri, S.P.; Rivera-Monroy, V.H.; Twilley, R.R. A comparison of nitrogen fixation (acetylene reduction) among three species of mangrove litter, sediments, and pneumatophores in south Florida, USA. *Hydrobiologia* **1997**, *356*, 73–79. [[CrossRef](#)]
141. Shiau, Y.-J.; Lin, M.-F.; Tan, C.-C.; Tian, G.; Chiu, C.-Y. Assessing N₂ fixation in estuarine mangrove soils. *Estuar. Coast. Shelf Sci.* **2017**, *189*, 84–89. [[CrossRef](#)]
142. Romero, I.C.; Jacobson, M.; Fuhrman, J.A.; Fogel, M.; Capone, D.G. Long-term nitrogen and phosphorus fertilization effects on N₂ fixation rates and *nifH* gene community patterns in mangrove sediments. *Mar. Ecol.* **2012**, *33*, 117–127. [[CrossRef](#)]
143. Vovides, A.G.; López-Portillo, J.; Bashan, Y. N₂-fixation along a gradient of long-term disturbance in tropical mangroves bordering the Gulf of Mexico. *Biol. Fertil. Soils* **2011**, *47*, 567–576. [[CrossRef](#)]
144. Zuberer, D.A.; Silver, W.S. Biological nitrogen fixation (acetylene reduction) associated with Florida mangroves. *Appl. Environ. Microbiol.* **1978**, *35*, 567–575. [[CrossRef](#)]
145. Sheridan, R.P. Epicaulous, nitrogen-fixing microepiphytes in a tropical mangal community, Guadeloupe, French West Indies. *Biotropica* **1991**, *23*, 530–541. [[CrossRef](#)]
146. Mann, F.D.; Steinke, T.D. Biological nitrogen fixation (acetylene reduction) associated with blue-green algal (cyanobacterial) communities in the Beachwood Mangrove Nature Reserve. I. The effect of environmental factors on acetylene reduction activity. *S. Afr. Tydskr. Plantk.* **1989**, *55*, 438–446. [[CrossRef](#)]
147. Mann, F.D.; Steinke, T.D. Biological nitrogen fixation (acetylene reduction) associated with blue-green algal (cyanobacterial) communities in the Beachwood Mangrove Nature Reserve. II. Seasonal variation in acetylene reduction activity. *S. Afr. J. Bot.* **1993**, *59*, 1–8.
148. Pelagri, S.P.; Twilley, R.R. Heterotrophic nitrogen fixation (acetylene reduction) during leaf-litter decomposition of two mangrove species from south Florida, USA. *Mar. Biol.* **1998**, *131*, 53–61.
149. Sengupta, A.; Chaudhuri, S. Ecology of heterotrophic dinitrogen fixation in the rhizosphere of mangrove plant community at the Ganges River estuary in India. *Oecologia* **1991**, *87*, 560–564. [[CrossRef](#)]
150. Vovides, A.G.; Bashan, Y.; López-Portillo, J.; Guevara, R. Nitrogen fixation in preserved, reforested, naturally regenerated and impaired mangroves as an indicator of functional restoration in mangroves in an arid region of Mexico. *Restor. Ecol.* **2010**, *19*, 236–244. [[CrossRef](#)]
151. Sheridan, R.P. Nitrogen fixation by epicaulous cyanobacteria in the Pointe de la Saline mangrove community, Guadeloupe, French West Indies. *Biotropica* **1992**, *24*, 571–574. [[CrossRef](#)]
152. Lugo-mela, C.; Bergman, B. Biological N₂-fixation on mangrove pneumatophores: Preliminary observations and perspectives. *Ambio* **2002**, *31*, 612–623. [[CrossRef](#)]
153. Potts, M.; Whitton, B.A. Nitrogen fixation by blue-green algal communities in the intertidal zone of the lagoon of Aldabra Atoll. *Oecologia* **1977**, *27*, 275–283. [[CrossRef](#)]
154. Potts, M. Nitrogen fixation (acetylene reduction) associated with communities of heterocystous and non-heterocystous blue-green algae in mangrove forests of Sinai. *Oecologia* **1979**, *39*, 359–373. [[CrossRef](#)] [[PubMed](#)]
155. Boto, K.G.; Robertson, A.I. The relationship between nitrogen fixation and tidal exports of nitrogen in a tropical mangrove system. *Estuar. Coast. Shelf Sci.* **1990**, *31*, 531–540. [[CrossRef](#)]

156. Woitchik, A.F.; Ohowa, B.; Kazungu, J.M.; Rao, R.G.; Goeyens, L.; Dehairs, F. Nitrogen enrichment during decomposition of mangrove leaf litter in an east African coastal lagoon (Kenya): Relative importance of biological nitrogen fixation. *Biogeochemistry* **1997**, *39*, 15–35. [[CrossRef](#)]
157. Uchino, F.; Hambali, G.G.; Yatazawa, M. Nitrogen-fixing bacteria from warty lenticellate bark of a mangrove tree, *Bruguiera gymnorrhiza* (L.) Lamk. *Appl. Environ. Microbiol.* **1984**, *47*, 44–48. [[CrossRef](#)]
158. Zuberer, D.A.; Silver, W.S. N₂-fixation (acetylene reduction) and the microbial colonization of mangrove roots. *New Phytol.* **1979**, *82*, 467–471. [[CrossRef](#)]
159. Toledo, G.; Bashan, Y.; Soeldner, A. Cyanobacteria and black mangroves in northwestern Mexico: Colonization, and diurnal and seasonal nitrogen fixation on aerial roots. *Can. J. Microbiol.* **1995**, *41*, 999–1011. [[CrossRef](#)]
160. Gotto, J.W.; Taylor, B.F. N₂ fixation associated with decaying leaves of the red mangrove (*Rhizophora mangle*). *Appl. Environ. Microbiol.* **1976**, *31*, 781–783. [[CrossRef](#)] [[PubMed](#)]
161. Qashqari, M.S.; Garcias-Bonet, N.; Fusi, M.; Booth, J.M.; Daffonchio, D.; Duarte, C.M. High temperature and crab density reduce atmospheric nitrogen fixation in Red Sea mangrove sediments. *Estuar. Coast. Shelf Sci.* **2020**, *232*, 106487. [[CrossRef](#)]
162. Capone, D.G. Determination of Nitrogenase Activity in Aquatic Samples Using the Acetylene Reduction Procedure. In *Aquatic Microbial Ecology*; Kemp, P.F., Sherr, B.F., Sherr, E.B., Cole, J.J., Eds.; Lewis Publishers: Boca Raton, FL, USA, 1993; pp. 621–632.
163. Wang, H.-T.; Su, J.-Q.; Zheng, T.-L.; Yang, X.-R. Impacts of vegetation, tidal process, and depth on the activities, abundances, and community compositions of denitrifiers in mangrove sediment. *Appl. Microbiol. Biotechnol.* **2014**, *98*, 9375–9387. [[CrossRef](#)] [[PubMed](#)]
164. Fernandes, S.O.; Loka Bharathi, P.A. Denitrification: An important pathway for nitrous oxide production in tropical mangrove sediments (Goa, India). *J. Environ. Qual.* **2010**, *39*, 1507–1516. [[CrossRef](#)]
165. Fernandes, S.O.; Loka Bharathi, P.A. Nitrate levels modulate denitrification activity in tropical mangrove sediments (Goa, India). *Environ. Monit. Assess.* **2011**, *173*, 117–125. [[CrossRef](#)]
166. Kristensen, E.; Mangion, P.; Tang, M.; Flindt, M.R.; Holmer, M.; Ulomi, S. Microbial carbon oxidation rates and pathways in sediments of two Tanzanian mangrove forests. *Biogeochemistry* **2011**, *103*, 143–158. [[CrossRef](#)]
167. Gao, G.-F.; Li, P.-F.; Zhong, J.-X.; Shen, Z.-J.; Chen, J.; Li, Y.-T.; Isabew, A.; Zhu, X.-Y.; Ding, Q.-S.; Zhang, S.; et al. *Spartina alterniflora* invasion alters soil bacterial communities and enhances soil N₂O emissions by stimulating soil denitrification in mangrove wetland. *Sci. Total Environ.* **2019**, *653*, 231–240. [[CrossRef](#)]
168. Pérez-Villalona, H.; Cornwell, J.C.; Ortiz-Zayas, J.R.; Cuevas, E. Sediment denitrification and nutrient fluxes in the San José Lagoon, a tropical lagoon in the highly urbanized San Juan Bay estuary, Puerto Rico. *Estuar. Coast.* **2015**, *38*, 2259–2278. [[CrossRef](#)]
169. Senthilkumar, B. Biogeochemical and Biophysical Aspects of Pichavarum Mangrove Ecosystem, South India. Ph.D. Thesis, Anna University, Chennai, India, 2007.
170. Rivera-Monroy, V.H.; Twilley, R.R.; Boustany, R.G.; Day, J.W.; Vera-Herrera, F.; del Carmen Ramirez, M. Direct denitrification in mangrove sediments in Terminos Lagoon, Mexico. *Mar. Ecol. Prog. Ser.* **1995**, *126*, 97–109. [[CrossRef](#)]
171. Guan, Q.S.; Cao, W.Z.; Wang, M.; Wu, G.J.; Wang, F.F.; Jiang, C.; Tao, Y.R.; Gao, Y. Nitrogen loss through anaerobic ammonium oxidation coupled with iron reduction in a mangrove wetland. *Eur. J. Soil Sci.* **2018**, *69*, 732–741. [[CrossRef](#)]
172. Zhang, M.; Dai, P.; Lin, X.; Lin, L.; Hetharua, B.; Zhang, Y.; Tian, Y. Nitrogen loss by anaerobic ammonium oxidation in a mangrove wetland of the Zhangjiang estuary, China. *Sci. Total Environ.* **2020**, *698*, 134291. [[CrossRef](#)]
173. Reis, C.R.G.; Nardoto, G.B.; Rochelle, A.L.C.; Vieira, S.A.; Oliveira, R.S. Nitrogen dynamics in subtropical fringe and basin mangrove forests inferred from stable isotopes. *Oecologia* **2017**, *183*, 841–848. [[CrossRef](#)]
174. Kristensen, E.; Andersen, F.Ø.; Holmboe, N.; Holmer, M.; Thongtham, N. Carbon and nitrogen mineralization in sediments of the Bangrong mangrove area, Phuket, Thailand. *Aquat. Microb. Ecol.* **2000**, *22*, 199–213. [[CrossRef](#)]
175. Nedwell, D.B.; Blackburn, T.H.; Wiebe, W.J. Dynamic nature of the turnover of organic carbon, nitrogen and sulphur in the sediments of a Jamaican mangrove forest. *Mar. Ecol. Prog. Ser.* **1994**, *110*, 223–231. [[CrossRef](#)]
176. Cao, W.; Guan, Q.; Li, Y.; Wang, M.; Liu, B. The contribution of denitrification and anaerobic ammonium oxidation to N₂ production in mangrove sediments of southeast China. *J. Soils Sediments* **2017**, *17*, 1767–1776. [[CrossRef](#)]

177. Krishnan, K.P.; Loka Bharathi, P.A. Organic carbon and iron modulate nitrification rates in mangrove swamps of Goa, south west coast of India. *Estuar. Coast. Shelf Sci.* **2009**, *84*, 419–426. [[CrossRef](#)]
178. Dunn, R.J.K.; Welsh, D.T.; Jordan, M.A.; Waltham, N.J.; Lemckert, C.J.; Teasdale, P.R. Benthic metabolism and nitrogen dynamics in a sub-tropical coastal lagoon: Microphytobenthos stimulate nitrification and nitrate reduction through photosynthetic oxygen evolution. *Estuar. Coast. Shelf Sci.* **2012**, *113*, 272–282. [[CrossRef](#)]
179. Xiao, K.; Wu, J.; Li, H.; Hong, Y.; Wilson, A.M.; Jiao, J.J.; Shanahan, M. Nitrogen fate in a subtropical mangrove swamp: Potential association with seawater-groundwater exchange. *Sci. Total Environ.* **2018**, *635*, 586–597. [[CrossRef](#)]
180. Amano, T.; Yoshinaga, I.; Yamagishi, T.; Van Thuoc, C.; The Thu, P.; Ueda, S.; Kato, K.; Sako, Y.; Suwa, Y. Contribution of anammox bacteria to benthic nitrogen cycling in a mangrove forest and shrimp ponds, Haiphong, Vietnam. *Microb. Environ.* **2011**, *26*, 1011040240. [[CrossRef](#)]
181. Purvaja, R.; Ramesh, R.; Ray, A.K.; Rixen, T. Nitrogen cycling: A review of the processes, transformations and fluxes in coastal ecosystems. *Curr. Sci.* **2008**, *94*, 1419–1438.
182. Luvizotto, D.M.; Araujo, J.E.; de Cássia, P.; Silva, M.; Dias, A.C.F.; Kraft, B.; Tegetmeyer, H.; Strous, M.; Andreote, F.D. The rates and players of denitrification, dissimilatory nitrate reduction to ammonia (DNRA) and anaerobic ammonia oxidation (anammox) in mangrove soils. *An. Acad. Bras. Cienc.* **2019**, *91*, e20180373. [[CrossRef](#)] [[PubMed](#)]
183. Dong, L.F.; Sobey, M.N.; Smith, C.J.; Rusmana, I.; Phillips, W.; Stott, A.; Osborn, A.M.; Nedwell, D.B. Dissimilatory reduction of nitrate to ammonium, not denitrification or anammox, dominates benthic nitrate reduction in tropical estuaries. *Limnol. Oceanogr.* **2011**, *56*, 279–291. [[CrossRef](#)]
184. Fernandes, S.O.; Michotey, V.D.; Guasco, S.; Bonin, P.C.; Loka Bharathi, P.A. Denitrification prevails over anammox in tropical mangrove sediments. *Mar. Environ. Res.* **2012**, *74*, 9–19. [[CrossRef](#)]
185. Zheng, Y.; Hou, L.; Zhang, Z.; Chen, F.; Gao, D.; Yin, G.; Han, P.; Dong, H.; Liang, X.; Yang, Y.; et al. Anaerobic ammonium oxidation (anammox) bacterial diversity, abundance, and activity in sediments of the Indus estuary. *Estuar. Coast. Shelf Sci.* **2020**, *243*, 106925. [[CrossRef](#)]
186. Meyer, R.L.; Risgaard-Petersen, N.; Allen, D.E. Correlation between anammox activity and microscale distribution of nitrite in a subtropical mangrove sediment. *Appl. Environ. Microbiol.* **2005**, *71*, 6142–6149. [[CrossRef](#)]
187. Cao, W.; Yang, J.; Li, Y.; Liu, B.; Wang, F.; Chang, C. Dissimilatory nitrate reduction to ammonium conserves nitrogen in anthropogenically affected subtropical mangrove sediments in southeast China. *Mar. Pollut. Bull.* **2016**, *110*, 155–161. [[CrossRef](#)]
188. Molnar, N.; Welsh, D.T.; Marchand, C.; Deborde, J.; Meziane, T. Impacts of shrimp farm effluent on water quality, benthic metabolism and N-dynamics in a mangrove forest (New Caledonia). *Estuar. Coast. Shelf Sci.* **2013**, *117*, 12–21. [[CrossRef](#)]
189. Dunn, R.J.K.; Welsh, D.T.; Jordan, M.A.; Teasdale, P.R.; Lemckert, C.J. Influence of natural amphipod (*Victoripisa australiensis*) (Chilton, 1923) population densities on benthic metabolism, nutrient fluxes, denitrification and DNRA in sub-tropical estuarine sediment. *Hydrobiologia* **2009**, *628*, 95–109. [[CrossRef](#)]
190. Fernandes, S.O.; Bonin, P.C.; Michotey, V.D.; Garcia, N.; Loka Bharathi, P.A. Nitrogen-limited mangrove ecosystems conserve N through dissimilatory nitrate reduction to ammonium. *Sci. Rep.* **2012**, *2*, 419. [[CrossRef](#)]
191. Iizumi, H. Soil nutrient dynamics. In *Workshop on Mangrove Ecosystem Dynamics*; Cragg, S., Polunin, N., Eds.; UNESCO: New Delhi, India, 1986; pp. 171–180.
192. Chen, R.; Twilley, R.R. Patterns of mangrove forest structure and soil nutrient dynamics along the Shark River estuary, Florida. *Estuaries* **1999**, *22*, 955–970. [[CrossRef](#)]
193. Chen, J.; Wu, F.-H.; Xiao, Q.; Yang, Z.-H.; Huang, S.-K.; Wang, J.; Wu, Y.-G.; Dong, X.-J.; Pei, Z.-P.; Zheng, H.-L. Diurnal variation in nitric oxide emission flux from a mangrove wetland in Zhangjiang River estuary, China. *Estuar. Coast. Shelf Sci.* **2010**, *90*, 212–220. [[CrossRef](#)]
194. Castillo, J.A.A.; Apan, A.A.; Maraseni, T.N.; Salmo, S.G., III. Soil greenhouse gas fluxes in tropical mangrove forests and in land uses on deforested mangrove lands. *Catena* **2017**, *159*, 60–69. [[CrossRef](#)]
195. Allen, D.; Dalal, R.C.; Rennenberg, H.; Schmidt, S. Seasonal variation in nitrous oxide and methane emissions from subtropical estuary and coastal mangrove sediments, Australia. *Plant Biol.* **2011**, *13*, 126–133. [[CrossRef](#)]

196. Padhy, S.R.; Bhattacharya, P.; Dash, P.K.; Reddy, C.S.; Chakraborty, A.; Pathak, H. Seasonal fluctuations in three mode of greenhouse gas emission in relation to soil labile carbon pools in degraded mangrove, Sundarban, India. *Sci. Total Environ.* **2020**, *705*, 135909. [[CrossRef](#)] [[PubMed](#)]
197. Chen, G.; Chen, B.; Yu, D.; Ye, Y.; Tam, N.F.Y.; Chen, S. Soil greenhouse gas emissions reduce the contribution of mangrove plants to the atmospheric cooling effect. *Environ. Res. Lett.* **2016**, *11*, 124019. [[CrossRef](#)]
198. Chauhan, R.; Datta, A.; Ramanathan, A.; Adhya, T.K. Factors influencing spatio-temporal variation of methane and nitrous oxide emission from a tropical mangrove of eastern coast of India. *Atmos. Environ.* **2015**, *107*, 95–106. [[CrossRef](#)]
199. Konnerup, D.; Betancourt-Portela, J.M.; Villamil, C.; Parra, J.P. Nitrous oxide and methane emissions from the restored mangrove ecosystem of the Ciénaga de Santa Marta, Colombia. *Estuar. Coast. Shelf Sci.* **2014**, *140*, 43–51. [[CrossRef](#)]
200. Chen, G.C.; Tam, N.F.Y.; Ye, Y. Summer fluxes of atmospheric greenhouse gases N₂O, CH₄ and CO₂ from mangrove soil in South China. *Sci. Total Environ.* **2010**, *408*, 2761–2767. [[CrossRef](#)]
201. Queiroz, H.M.; Artur, A.G.; Taniguchi, C.A.K.; da Silva, M.R.S.; do Nascimento, J.C.; Nóbrega, G.N.; Otero, X.L.; Ferreira, T.O. Hidden contribution of shrimp farming effluents to greenhouse gas emissions from mangrove soils. *Estuar. Coast. Shelf Sci.* **2019**, *221*, 8–14. [[CrossRef](#)]
202. Chen, G.C.; Tam, N.F.Y.; Ye, Y. Spatial and seasonal variations of atmospheric N₂O and CO₂ fluxes from a subtropical mangrove swamp and their relationships with soil characteristics. *Soil Biol. Biochem.* **2012**, *48*, 175–181. [[CrossRef](#)]
203. Wang, H.; Liao, G.; D'Souza, M.; Yu, X.; Yang, J.; Yang, X.; Zheng, T. Temporal and spatial variations of greenhouse gas fluxes from a tidal mangrove wetland in southeast China. *Environ. Sci. Pollut. Res.* **2016**, *23*, 1873–1885. [[CrossRef](#)] [[PubMed](#)]
204. Chen, G.C.; Ulumuddin, Y.I.; Pramudji, S.; Chen, S.Y.; Chen, B.; Ye, Y.; Ou, D.Y.; Ma, Z.Y.; Huang, H.; Wang, J.K. Rich carbon and nitrogen but low atmospheric greenhouse gas fluxes from North Sulawesi mangrove swamps in Indonesia. *Sci. Total Environ.* **2014**, *487*, 91–96. [[CrossRef](#)] [[PubMed](#)]
205. Chauhan, R.; Ramanathan, A.L.; Adhya, T.K. Assessment of methane and nitrous oxide flux from mangroves along the east coast of India. *Geofluids* **2008**, *8*, 321–332. [[CrossRef](#)]
206. Bauza, J.F.; Morell, J.M.; Corredor, J.E. Biogeochemistry of nitrous oxide production in the red mangrove (*Rhizophora mangle*) forest sediments. *Estuar. Coast. Shelf Sci.* **2002**, *55*, 697–704. [[CrossRef](#)]
207. Allen, D.E.; Dalal, R.C.; Rennenberg, H.; Meyer, R.L.; Reeves, S.; Schmidt, S. Spatial and temporal variation of nitrous oxide and methane flux between subtropical mangrove sediments and the atmosphere. *Soil Biol. Biochem.* **2007**, *39*, 622–631. [[CrossRef](#)]
208. Muñoz-Hincapié, M.; Morell, J.M.; Corredor, J.E. Increase of nitrous oxide flux to the atmosphere upon nitrogen addition to red mangroves sediments. *Mar. Pollut. Bull.* **2002**, *44*, 992–996. [[CrossRef](#)]
209. Cameron, C.; Hutley, L.B.; Friess, D.A.; Brown, B. High greenhouse gas emissions mitigation benefits from mangrove rehabilitation in Sulawesi, Indonesia. *Ecosyst. Serv.* **2019**, *40*, 101035. [[CrossRef](#)]
210. Chen, G.; Chen, J.; Ou, D.; Tam, N.F.T.Y.; Chen, S.; Zhang, Q.; Chen, B.; Ye, Y. Increased nitrous oxide emissions from intertidal soil receiving wastewater from dredging shrimp pond sediments. *Environ. Res. Lett.* **2020**, *15*, 094015. [[CrossRef](#)]
211. Krithika, K.; Purjaya, R.; Ramesh, R. Fluxes of methane and nitrous oxide from an Indian mangrove. *Curr. Sci.* **2008**, *94*, 218–224.
212. Müller, D.; Bange, H.W.; Warneke, T.; Rixen, T.; Müller, M.; Mujahid, A.; Notholt, J. Nitrous oxide and methane in two tropical estuaries in a peat-dominated region of northwestern Borneo. *Biogeosciences* **2016**, *13*, 2415–2428. [[CrossRef](#)]
213. Bange, H.W.; Sim, C.H.; Bastian, D.; Kallert, J.; Kock, A.; Mujahid, A.; Müller, M. Nitrous oxide (N₂O) and methane (CH₄) in rivers and estuaries of northwestern Borneo. *Biogeosciences* **2019**, *16*, 4321–4335. [[CrossRef](#)]
214. Barnes, J.; Ramesh, R.; Purvaja, R.; Rajkumar, A.N.; Sentil Kumar, B.; Krithika, K.; Ravichandran, K.; Uher, G.; Upstill-Goddard, R. Tidal dynamics and rainfall control N₂O and CH₄ emissions from a pristine mangrove creek. *Geophys. Res. Lett.* **2006**, *33*, L15405. [[CrossRef](#)]
215. Maher, D.T.; Sippo, J.Z.; Tait, D.R.; Holloway, C.; Santos, I.R. Pristine mangrove creek waters are a sink of nitrous oxide. *Sci. Rep.* **2016**, *6*, 25701. [[CrossRef](#)] [[PubMed](#)]
216. Divia, J.I.; Neetha, V.; Hariharan, G.; Kakolee, B.; Purvaja, V.; Ramesh, R. N₂O flux from South Andaman mangroves and surrounding creek waters. *Int. J. Ocean. Oceanogr.* **2013**, *7*, 73–82.

217. Murray, R.; Erler, D.; Rosentreter, J.; Maher, D.; Eyre, B. A seasonal source and sink of nitrous oxide in mangroves: Insights from concentration, isotope, and isotopomer measurements. *Geochim. Cosmochim. Acta* **2018**, *238*, 169–192. [[CrossRef](#)]
218. Reading, M.J.; Santos, I.R.; Maher, D.T.; Jeffrey, L.C.; Tait, D.R. Shifting nitrous oxide source/sink behaviour in a subtropical estuary revealed by automated time series observations. *Estuar. Coast. Shelf Sci.* **2017**, *194*, 66–76. [[CrossRef](#)]
219. Sturm, K.; werner, U.; Grinham, A.; Yuan, Z. tidal variability in methane and nitrous oxide emissions along a subtropical estuarine gradient. *Estuar. Coast. Shelf Sci.* **2017**, *192*, 159–169. [[CrossRef](#)]
220. Rao, G.D.; Sarma, V.V.S.S. Contribution of N₂O emissions to the atmosphere from Indian monsoonal estuaries. *Tellus B* **2013**, *65*, 19660. [[CrossRef](#)]
221. Murray, R.; Erler, D.; Rosentreter, J.; Wells, N.; Eyre, B. Seasonal and spatial controls on N₂O concentrations and emissions in low-nitrogen estuaries: Evidence from three tropical systems. *Mar. Chem.* **2020**, *221*, 103779. [[CrossRef](#)]
222. Chen, N.; Wu, J.; Zhou, X.; Chen, Z.; Lu, T. Riverine N₂O production, emissions and export from a region dominated by agriculture in Southeast Asia (Jiulong River). *Agric. Ecosyst. Environ.* **2015**, *208*, 37–47. [[CrossRef](#)]
223. Lin, H.; Dai, M.; Kao, S.J.; Wang, L.; Roberts, E.; Yang, J.Y.T.; Huang, T.; He, B. Spatiotemporal variability of nitrous oxide in a large eutrophic estuarine system: The Pearl River estuary, China. *Mar. Chem.* **2016**, *182*, 14–24. [[CrossRef](#)]
224. Liu, X.L.; Bai, L.; Wang, Z.L.; Li, J.; Yue, F.J.; Li, S.L. Nitrous oxide emissions from river network with variable nitrogen loading in Tianjin, China. *J. Geochem. Expl.* **2015**, *157*, 153–161. [[CrossRef](#)]
225. Wells, N.S.; Maher, D.T.; Erlet, D.V.; Hipsey, M.; Rosentreter, J.A.; Eyre, B.D. Estuaries as sources and sinks of N₂O across a land-use gradient in subtropical Australia. *Glob. Biogeochem. Cycle.* **2018**, *32*, 877–894. [[CrossRef](#)]
226. Manjrekar, S.; Uskaikar, H.; Morajkar, S. Seasonal production of nitrous oxide in a tropical estuary off western India. *Reg. Stud. Mar. Sci.* **2020**, *39*, 101418. [[CrossRef](#)]
227. Mohammed, S.M.; Johnstone, R.W. Porewater profiles and nutrient sediment-water exchange in a tropical mangrove waterway, Mapopwe Creek, Chwaka Bay, Zanzibar. *Afr. J. Ecol.* **2002**, *40*, 172–178. [[CrossRef](#)]
228. Jiang, S.; Müller, M.; Jin, J.; Wy, Y.; Zhu, K.; Zhang, G.; Mujahid, A.; Rixen, T.; Fakharuddin Muhamad, M.; Sien Aun Sia, E.; et al. Dissolved inorganic nitrogen in a tropical estuary in Malaysia: Transport and transformation. *Biogeosciences* **2019**, *16*, 2821–2836. [[CrossRef](#)]
229. Terada, K.; Koibuchi, Y.; Isobe, M. Rainfall effect on sediment and nutrient fluxes in a small mangrove river, Okinawa, Japan. *J. Mar. Res.* **2017**, *75*, 1–17. [[CrossRef](#)]
230. Davis, S.E., III; Lirman, D.; Wozniak, J.R. Nitrogen and Phosphorus Exchange among Tropical Coastal Ecosystems. In *Ecological Connectivity among Tropical Coastal Ecosystems*; Nagelkerken, I., Ed.; Springer: Amsterdam, The Netherlands, 2009; pp. 9–43.
231. Davis, S.E., III; Childers, D.L.; Day, J.W., Jr.; Rudnick, D.T.; Sklar, F.H. Wetland-water column exchanges of carbon, nitrogen, and phosphorus in a Southern Everglades dwarf mangrove. *Estuaries* **2001**, *24*, 610–622. [[CrossRef](#)]
232. Boto, K.G.; Bunt, J.S. Tidal export of particulate organic matter from a northern Australian mangrove system. *Estuar. Coast. Shelf Sci.* **1981**, *13*, 247–255. [[CrossRef](#)]
233. Rivera-Monroy, V.H.; Day, J.W.; Twilley, R.R.; Vera-Herrera, F.; Coronado-Molina, C. Flux of nitrogen and sediment in a fringe mangrove forest in Terminos Lagoon, Mexico. *Estuar. Coast. Shelf Sci.* **1995**, *40*, 139–160. [[CrossRef](#)]
234. Davis, S.E., III; Childers, D.L.; Day, J.W., Jr.; Rudnick, D.T.; Sklar, F.H. Nutrient dynamics in vegetated and unvegetated areas of a Southern Everglades mangrove creek. *Estuar. Coast. Shelf Sci.* **2001**, *52*, 753–768. [[CrossRef](#)]
235. Barboza, C.D.N.; Paes, E.T.; de Andrade Jandre, K.; Marques, A.N., Jr. Concentrations and fluxes of nutrients and suspended organic matter in a tropical estuarine system: The Tinharé-Boipeba Islands Archipelago (Baixo Sul Baiano, Brazil). *J. Coast. Res.* **2014**, *30*, 1197–1209. [[CrossRef](#)]
236. Tay, H.W.; Bryan, K.R.; Pilditch, C.A.; Park, S.; Hamilton, D.P. Variations in nutrient concentrations at different time scales in two shallow tidally dominated estuaries. *Mar. Freshw. Res.* **2012**, *63*, 95–109. [[CrossRef](#)]

237. Valiela, I.; Elmstrom, E.; Lloret, J.; Stone, T.; Camilli, L. Tropical land-sea couplings: Role of watershed deforestation, mangrove estuary processing, and marine inputs on N fluxes in coastal Pacific Panama. *Sci. Total Environ.* **2018**, *630*, 126–140. [[CrossRef](#)] [[PubMed](#)]
238. Rivera-Monroy, V.H.; Twilley, R.R.; Davis, S.E., III; Childers, D.L.; Simard, M.; Chambers, R.; Jaffe, R.; Boyer, J.N.; Rudnick, D.T.; Zhang, K.; et al. The role of the Everglades mangrove ecotone region (EMER) in regulating nutrient cycling and wetland productivity in South Florida. *Crit. Rev. Environ. Sci. Technol.* **2011**, *41* (Suppl. S1), 633–669. [[CrossRef](#)]
239. Ray, R.; Majumder, N.; Das, S.; Chowdhury, C.; Kumar Jana, T. Biogeochemical cycle of nitrogen in a tropical mangrove ecosystem, east coast of India. *Mar. Chem.* **2014**, *167*, 33–43. [[CrossRef](#)]
240. Adame, M.F.; Virdis, B.; Lovelock, C.E. Effect of geomorphological setting and rainfall on nutrient exchange in mangroves during tidal inundation. *Mar. Freshw. Res.* **2010**, *61*, 1197–1206. [[CrossRef](#)]
241. Adame, M.F.; Lovelock, C.E. Carbon and nutrient exchange of mangrove forests with the coastal ocean. *Hydrobiologia* **2011**, *663*, 23–50. [[CrossRef](#)]
242. Hamilton, S.E.; Casey, D. Creation of a high spatio-temporal resolution global database of continuous mangrove forest cover for the 21st century. *Glob. Ecol. Biogeogr.* **2016**, *25*, 729–738. [[CrossRef](#)]
243. Alongi, D.M. Carbon cycling in the world's mangrove ecosystems revisited: Significance of non-steady state diagenesis and subsurface linkages between the forest floor and the coastal ocean. *Forests* **2020**, *11*, 977. [[CrossRef](#)]
244. Hillebrand, H.; Sommer, U. The nutrient stoichiometry of benthic microalgal growth: Redfield proportions are optimal. *Limnol. Oceanogr.* **1999**, *44*, 440–446. [[CrossRef](#)]
245. Cerón, R.M.; Cerón, J.G.; Muriel, M.; Anguebes, F.; Ramirez, M.; Zavala, J.; Carballo, C.; Escoffie, R.C. Spatial and Temporal Distribution of Throughfall Deposition of Nitrogen and Sulfur in the Mangrove Forests Associated with Terminos Lagoon. In *Current Air Quality Issues*; Nejadkoorki, F., Ed.; Intech Open Ltd: London, UK, 2015; pp. 147–164.
246. Seitzinger, S.; Harrison, J.A.; Böhlke, J.K.; Bouwman, A.F.; Lowrance, R.; Peterson, B.; Tobias, C.; Van Drecht, G. Denitrification across landscapes and waterscapes: A synthesis. *Ecol. Appl.* **2006**, *16*, 2064–2090. [[CrossRef](#)]
247. Reis, C.R.; Nardoto, G.B.; Oliveira, R.S. Global overview on nitrogen dynamics in mangroves and consequences of increasing nitrogen availability for these systems. *Plant Soil* **2017**, *410*, 1–9. [[CrossRef](#)]
248. Alvarez-Cobelas, M.; Angeler, D.G.; Sánchez-Carrillo, S. Export of nitrogen from catchments: A worldwide analysis. *Environ. Pollut.* **2008**, *156*, 261–269. [[CrossRef](#)] [[PubMed](#)]
249. Beusen, A.H.W.; Bouwman, A.F.; Van Beck, L.P.H.; Mogollón, J.M.; Middelburg, J.J. Global riverine N and P transport to ocean increased during the 20th century despite increased retention along the aquatic continuum. *Biogeosciences* **2016**, *13*, 2441–2451. [[CrossRef](#)]
250. Fowler, D.; Coyle, M.; Skiba, U.; Sutton, M.A.; Cape, J.N.; Reis, S.; Sheppard, L.J.; Jenkins, A.; Grizzetti, B.; Galloway, J.N.; et al. The global nitrogen cycle in the twenty-first century. *Philos. Trans. R. Soc. B* **2013**, *368*, 20130164. [[CrossRef](#)]
251. Alongi, D.M. Carbon balance in salt marsh and mangrove ecosystems: A global synthesis. *J. Mar. Sci. Eng.* **2020**, *8*, 767. [[CrossRef](#)]

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